



A ROLE OF P-GLYCOPROTEIN IN MODULATION OF ANTIBIOTIC PHARMACOKINETICS

A. Haritova

Department of Pharmacology, Physiology of Animals and Physiological Chemistry, Faculty of Veterinary Medicine, Trakia University, 6000 Stara Zagora, Bulgaria

ABSTRACT

P-glycoprotein (P-gp) belongs to superfamily of ABC (ATP-binding cassette) drug transporters. It is expressed in intestines, brain, kidneys, testes, liver, adrenal gland, lungs heart and eyes. This protein functions as a biological barrier by extruding toxic substances and xenobiotics out of cells. P-gp could modulate pharmacokinetics of antibacterial drugs through limitation of their oral absorption and penetration into target organs. Drug-drug interactions with antibacterials could be mediated by inhibition or induction of P-gp. Among the antibiotics recognized as substrates and modulators of P-gp are structurally unrelated compounds as fluoroquinolones, macrolides, ansamycines, tetracyclines and antracyclines. These findings provide a basis for the understanding of the pharmacokinetics, pharmacodynamics and toxicodynamics of the antibiotics in healthy and diseased individuals.

Key words: P-glycoprotein, Antibiotics, Pharmacokinetics

INTRODUCTION

P-glycoprotein (P-gp) belongs to superfamily of ABC (ATP-binding cassette) drug transporters and it was initially identified because of its over expression in cultured tumor cells associated with an acquired cross-resistance to multiple cytotoxic anticancer agents (12, 14). It was also recognized to be expressed in many normal tissues, suggestive of a physiological function. P-gp is located in the apical membrane of the enterocyte of the gastrointestinal tract, suggesting that the transporter functions to facilitate excretion of substrates from the systemic circulation into the gut lumen. Similarly, localization of P-gp in the canalicular domain of the hepatocyte and the brush border of the proximal renal tubule is consistent with a role for the transporter in the biliary and urinary excretion of xenobiotics and endogenous substrates. In addition, it has a protective function at the site of important physiological barriers such as blood-brain and blood-testes. It is also

expressed in tissues as adrenal gland, lungs, heart and eyes (43).

P-gp shows broad substrate specificity, recognizing a large number of compounds with unrelated pharmacological properties. It acts as one of general means for cells to protect themselves from undesirable invasion by compounds, including antibiotics, which freely diffuse across membranes. Function of this protein affects drug pharmacokinetics through limitation of their oral absorption and penetration into target organs such as the brain and testes (18, 26). In individuals with increased P-gp expression or function, reduced oral bioavailability, decreased maximal plasma concentrations, increased renal clearance and reduced area under the curve (AUC) would be expected (15).

This work is aimed to discuss the role of P-gp in the absorption and disposition of antibiotics and participation of these drugs in drug-drug interactions by modulation of the activity of P-gp.

Antibacterial drugs

The role of P-gp in the modulation of antibiotic pharmacokinetics has been mainly studied for fluoroquinolones and macrolides. Nowadays the number of evidences about the interaction of P-gp with other antibacterial

Correspondence to: Aneliya Haritova, Department of Pharmacology, Physiology of Animals and Physiological Chemistry, Faculty of Veterinary Medicine, Trakia University, 6000 Stara Zagora, Bulgaria, Phone + 35942699622, E-mail: aneliah@sz.inetg.bg

drugs is significantly increasing.

***β*-lactams**

Transporters, including P-gp, identified at the blood–brain and blood–CSF barriers are important for the disposition of *β*-lactams because active efflux may be detrimental for treatment of meningitis and other infections of the CNS (27, 34). For example, failures in treatment of inflammation of CNS with cefalothin have been attributed to the active efflux of this molecule (35). Dicloxacillin is recognized as a substrate for P-gp and inter-individual differences in its pharmacokinetics were explained by variations in MDR1 genotype (22). These findings are supported by the fact that the induction of this protein by rifampin administration provokes decreasing of differences in the dicloxacillin pharmacokinetics (clearance, AUC and C_{max}) between various MDR1 genotypes (22). Isoxazolyl-penicillin flucloxacillin has the potential to induce expression of both CYP3A4 as well as P-gp, through activation of the nuclear hormone receptor pregnane-X-receptor (PXR) which would offer an explanation for the observed clinical drug–drug interactions between the antibiotic and cyclosporine (11).

Fluoroquinolones

Fluoroquinolones have been reported as a class of drugs able to undergo efflux, which can explain the low bioavailability of some of them after oral administration. It has been shown in *in situ* and *in vitro* studies, that absorption of grepafloxacin and sparfloxacin is increased by concomitant administration of P-gp inhibitors (23). It has to be taken into account that involvement of other transporters in addition to P-gp can also play a role because there are not highly specific compounds with inhibitory activity. For example, in ciprofloxacin transport are involved more than one efflux and even influx proteins (23). Fluoroquinolones are not only substrates but also inhibitors for ABC transporters. Grepafloxacin, levofloxacin and sparfloxacin inhibit P-gp activity in MDCKII-MDR1 and Caco2 cell lines which is a prerequisite for modulated oral absorption and disposition of other substrates for P-gp as macrolide drugs when administered concomitantly (33).

Inhibition of P-gp contributes to cellular accumulation of ciprofloxacin in macrophages but mainly in the cytosolic compartment, which can partly assist in elimination of intracellular bacteria as *Listeria*

monocytogenes and *Staphylococcus aureus* (32). Between fluoroquinolones licensed for use in animals, danofloxacin mesylate appears to be a substrate for P-gp and MRP2 in Caco2 cell lines. Its absorption and secretion are decreased by simultaneously applied ciprofloxacin (29). Other studies with chicken lymphocytes show that danofloxacin base and danofloxacin mesylate at high concentrations, which could be reached in poultry after administration of usual dosage regimens, have inhibitory effect mainly on function of P-gp and to a much lesser extent on breast cancer resistance protein (BCRP) (10).

Brain distribution of fluoroquinolones is regulated by P-gp and a parallel has been observed between the propensity of fluoroquinolones to induce seizures and their rate of efflux from the CNS (4). Concentrations of grepafloxacin and a new fluoroquinolone drug HSR-903 are much higher in the brain in *mdr1a* (–/–) knockout mice (37). Moreover, grepafloxacin, sparfloxacin, and norfloxacin, were strong inhibitors of efflux of HSR-903, although other quinolone derivatives, which are relatively hydrophilic, had less effect on HSR-903 uptake by the brain capillary endothelial cells, suggesting a competition for the efflux transporter proteins, including P-gp (19).

Clearance of fluoroquinolones can be accelerated by the action of MDR1 in the intestine, kidney, liver or CNS (24). Lower total body clearance of grepafloxacin in *mdr1a* (–/–) and *mdr1a/1b* (–/–) suggests that urinary and biliary excretion, intestinal secretion and metabolism are reduced in these mice (25).

P-gp could affect pharmacokinetics of fluoroquinolones in different way in healthy and diseased animals. In rats with hepatic fibrosis decreased bioavailability and increased elimination of orally administered ofloxacin was observed due to up-regulation of P-gp and increased activities of cytochrome P₄₅₀ (CYP₄₅₀) in small intestines (40).

Macrolide antibiotics

It is well known that macrolides are antibiotics that have inhibitory activity on metabolizing enzymes. After discovery of ABC transport proteins it is clear that they also interact with P-gp. Their clearance can be accelerated by the action of MDR1 in the intestine, kidney, liver or CNS (36). Erythromycin exhibited significant efflux out of MDCK-MDR1 cells transfected with the human *mdr1* gene, suggesting that erythromycin is a good substrate for P-gp.

Intestinal P-gp determines oral absorption and disposition of erythromycin leading to increased plasma and tissue levels in Mdr1a (-/-) mice (30). It was demonstrated in *in vivo* experiments with New Zealand albino (New Zealand White) rabbits that P-gp restricts topical erythromycin absorption across the cornea. Therefore, ocular bioavailability of P-gp substrates can be significantly enhanced by proper selection of P-gp inhibitors (5). Inhibition of P-gp contributes to accumulation of azitromycin in macrophages and thus to elimination of intracellular bacteria (32).

Erythromycin is known with its inhibitory activity on the enzymes from CYP₄₅₀ family and P-gp function (31). For instance, mechanism of the pharmacokinetic interaction between oral ximelagatran (an oral direct thrombin inhibitor) and erythromycin involve inhibition of transport proteins, possibly P-gp, resulting in decreased biliary excretion and increased bioavailability of melagatran (6). Clarithromycin reduces the renal secretion of digoxin by blocking P-gp activity in the renal tubule (39).

At the same time, efflux by P-gp does not have significant influence in the kinetics of relatively new macrolides telithromycin and roxithromycin (21).

Ansamycines

Rifampicin is characterized as an inducer of both CYP₄₅₀ and MDR expression and it can reduce the blood level of several drugs by induction of these proteins. Since cyclosporine is a substrate and rifampicin is an inducer for both CYP3A4 and P-gp, the observed increased clearance and decreased bioavailability of the immunosuppressive agent during rifampicin treatment are most probably due to a combination of CYP3A4 and P-gp induction (38). Rifampicin can also decrease the extent of absorption of other P-gp substrates as ranitidine by induction of the activity of this protein (16).

Species specific differences were observed in the effect of rifampicin on transcriptional level of expression of MDR1. It upregulates MDR1 mRNA expression in human hepatocytes and hepatoma cells but in rats Mdr1a and Mdr1b are not inducible by this antibiotic (20).

Tetracyclines

Oxytetracycline has been used for decennia in veterinary medicine due to its extensive spectrum of antimicrobial activity. A major limitation has been, and still remains, its low bioavailability following oral administration.

Experiments with Caco-2 cells show that PSC833, a potent inhibitor of P-gp, decreased the secretion of oxytetracycline without affecting its absorption and indicate that this drug is a substrate for this protein. The affinity of oxytetracycline to this transporter seems to be rather low, as suggested by the low efflux ratio of 1:1.3. In competition experiments, oxytetracycline decreased the effluxes of other P-gp substrates such as Rhodamine-123 and ivermectin, findings of clinical relevance, as they clearly indicate potential drug-drug interactions at the level of P-gp-mediated drug transport (28).

Doxycycline, a tetracycline antimicrobial agent with structural similarity to doxorubicin, induced expression of MDR1 mRNA and P-gp. It reduced intracellular accumulation of doxorubicin, a substrate for P-gp and thus could contribute to clinical chemotherapeutic failure in cancer patients as a consequence of generating P-gp-expressing. Moreover, cells expressing P-gp demonstrated reduced intracellular accumulation of this tetracycline compared to that of cells that did not express P-gp suggesting that doxycycline is a substrate for P-gp (17).

Jonofore and other antibiotics

Dietary antibiotic as monensin, a model for dietary toxin, altered P-gp expression in poultry. Monensin increased P-gp expression in the liver and duodenum. Other antibiotic, bacitracin, reduced P-gp expression by 45% in the liver, but did not alter expression in the duodenum. This study indicates that dietary constituents regulate the expression of P-gp and these changes may represent an important physiological response to foods containing toxins (1).

Anticancer antibiotics

Nowadays, treatment of cancer in small animals became more and more important in veterinary practice. Multidrug resistance in cancer cells is discussed widely during last decade and a lot of efforts were spent to find a way to overcome this complicated problem (8). This section is pointed at the problems with multidrug resistance associated with the use of anticancer antibiotics and mediated by P-gp, only. Doxorubicin is one of the classical examples for an anticancer agent which is good substrate for P-gp. In P-gp-overexpressing cells, it becomes a difficult task to maintain a high intracellular doxorubicin level for a reasonable length of time (42). The pharmacokinetics of doxorubicin in blood and tissues of female

Balb/c mice was altered by earlier exposure to doxorubicin, as the animals that were treated once a week for 2 weeks showed an increased rate of doxorubicin elimination from blood and tissues following the second treatment because of overexpression of P-gp. These results have implications both in multiple-dosing regimens, as well as multiple-drug regimens, where doxorubicin is used in combination with other drugs that are substrates for P-gp (9). Idarubicin, an anthracycline antineoplastic agent, was characterized as a substrate for P-gp and its uptake in the cells is increased by verapamil and amiodarone, which function as inhibitors of this protein. Thus, these drug combinations increase acute negative inotropic effect of anthracyclines on the heart (41).

CONCLUSION

We have to acknowledge that P-gp expression and function at apical site of the cell membranes could have significant influence on drug disposition by restricting the penetration of antibiotics in some tissues or limit their absorption from gastrointestinal tract or enhanced their elimination. Opposite effect could be observed if this protein is down-regulated or inhibited. Interplay with drug metabolism also has to be taken into account (2). Notwithstanding, real situations became more complicated because other transporters as BCRP and MRPs were recognized as proteins which can modulate drug kinetics. Moreover, there is substantial overlap in substrate specificities for the different ABC transporters. Transport proteins which function at the basal site of cell membranes belonging to solute carrier family of transporters are also involved. Species specific differences in the expression pattern and activity of drug transporters require their investigation in different animal species and even breeds (7, 15). In dose determination strategy it has to be considered that efflux transport is energy dependent and saturable process when high concentrations of drugs are administered. In addition, bacterial cells also have ABC and drug efflux transporters which function in a coupled exchange with protons or sodium ions along a concentration gradient as symport or antiport (3, 13). Therefore, it is important to evaluate the role of drug transporters in target animal species not only from pharmacokinetic point of view but also because of pharmacodynamic and toxicological considerations.

REFERENCES

1. Barnes, D.M., Expression of P-glycoprotein in the chicken. *Comp Biochem Physiol A*, 130:301–310, 2001.
2. Benet, L.Z., Cummins, C.L. and Wu, C.Y., Unmasking the dynamic interplay between efflux transporters and metabolic enzymes. *Int J Pharm*, 277:3–9, 2004.
3. Burnie, J.P., Maatthews, R.C., Carter, T., Beaulieu, E., Donohoe, M., Chapman, C., Williamson, P. and Hodgetts, S.J., Identification of an immunodominant ABC transporter in methicillin-resistant *Staphylococcus aureus* infections. *Infect Immun*, 68:3200–3209, 2000.
4. De Sarro, A., Cecchetti, V., Fravolini, V., Naccari, F., Tabarrini, O. and De Sarro, G., Effects of novel 6-desfluoroquinolones and classic quinolones on pentylenetetrazole-induced seizures in mice. *Antimicrob Agents Chemother*, 43:1729–1736, 1999.
5. Dey, S., Gunda, S. and Mitra, A.K., Pharmacokinetics of erythromycin in rabbit corneas after single-dose infusion: role of P-glycoprotein as a barrier to in vivo ocular drug absorption. *J Pharmacol Exp Ther*, 311:246–255, 2004.
6. Eriksson, U.G., Dorani, H., Karlsson, J., Fritsch, H., Hoffmann, K.J., Olsson, L., Sarich, T.C., Wall, U. and Schützer, K.M., Influence of erythromycin on the pharmacokinetics of ximelagatran may involve inhibition of P-glycoprotein-mediated excretion. *Drug Metab Dispos*, 34:775–782, 2006.
7. Geyer, J., Klintzsch, S., Meerkamp, K., Wohlke, A., Distl, O., Moritz, A. and Petzinger, E., Detection of the nt230(del4) MDR1 Mutation in White Swiss Shepherd Dogs: case reports of doramectin toxicosis, breed predisposition, and microsatellite analysis. *J Vet Pharmacol Ther*, 30:482–485, 2007.
8. Gottesman, M.M., Mechanism of cancer drug resistance. *Annu Rev Med*, 53: 615–627, 2002.
9. Gustafson, D.L. and Long, M.E., Alterations in P-glycoprotein expression in mouse tissues by

- doxorubicin: implications for pharmacokinetics in multiple dosing regimens. *Chemico-Biological Interactions*, 138:43–57, 2001.
10. Haritova, A., Schrickx, J. and Fink-Gremmels, J., Functional studies on the activity of efflux transporters in an *ex vivo* model with chicken splenocytes and evaluation of selected fluoroquinolones in this model. *Biochem Pharmacol*, 73:752–759, 2007.
 11. Huwyler, J., Wright, M.B., Gutmann, H. and Drewe, J., Induction of cytochrome P450 3A4 and P-glycoprotein by the isoxazolylpenicillin antibiotic flucloxacillin. *Curr Drug Metab*, 7:119–126, 2006.
 12. Juliano, R.L. and Ling, V., A surface glycoprotein modulating drug permeability in Chinese hamster ovary cell mutants. *Biochim Biophys Acta*, 455:152-162, 1976.
 13. Lage, H., ABC-transporters: implications on drug resistance from microorganisms to human cancers. *Int J Antimicrob Agents*, 22:188-199, 2003.
 14. Leonard, G.D., Fojo, T. and Bates, S.E. The role of ABC transporters in clinical practice. *Oncologist*, 8:411–424, 2003.
 15. Lin, J.H. and Yamazaki, M., Role of P-Glycoprotein in pharmacokinetics clinical implications. *Clin Pharmacokinet*, 42:59–98, 2003.
 16. Machavaram, K.K., Gundu, J. and Yamsani, M.R., Effect of ketoconazole and rifampicin on the pharmacokinetics of ranitidine in healthy human volunteers: a possible role of P-glycoprotein. *Drug Metabol Drug Interact*, 22:47–65, 2006.
 17. Mealey, K.L., Barhoumi, R., Burghardt, R.C., Safe, S. and Kochevar, D.T., Doxycycline induces expression of P glycoprotein in MCF-7 breast carcinoma cells. *Antimicrob Agents Chemother*, 46:755–761, 2002.
 18. Melaine, N., Lienard, M.-O., Dorval, I., Le Goascogne, C., Lejeune, H. and Jegou, B., Multidrug resistance genes and P-Glycoprotein in the testis of the rat, mouse, Guinea pig, and human. *Biol Reprod*, 67:1699–1707, 2002.
 19. Murata, M., Tamai, I., Kato, H., Nagata, O., Kato, H. and Tsuji, A., Efflux transport of a new quinolone antibacterial agent, HSR-903, across the blood–brain barrier. *J Pharmacol Exp Ther*, 290:51–57, 1999.
 20. Nishimura, M., Koeda, A., Suzuki, E., Kawano, Y., Nakayama, M., Satoh, T., Narimatsu, S. and Naito, S., Regulation of mRNA expression of MDR1, MRP1, MRP2 and MRP3 by prototypical microsomal enzyme inducers in primary cultures of human and rat hepatocytes. *Drug Metab Pharmacokinet*, 21:297–307, 2006.
 21. Pachot, J.I., Botham, R.P. and Haegele, K.D., Experimental estimation of the role of P-Glycoprotein in the pharmacokinetic behaviour of telithromycin, a novel ketolide, in comparison with roxithromycin and other macrolides using the Caco-2 cell model. *J Pharm Pharmaceut Sci*, 6:1–12, 2003.
 22. Putnam, W.S., Woo, J.M., Huang, Y. and Benet, L.Z., Effect of the MDR1 C3435T variant and P-glycoprotein induction on dicloxacillin pharmacokinetics. *J Clin Pharmacol*, 45:411–421, 2005.
 23. Rodriguez-Ibanez, M., Nalda-Molina, R., Montalar-Montero, M., Bermejo, M.V., Merino, V. and Garrigues, T.M., Transintestinal secretion of ciprofloxacin, grepafloxacin and sparfloxacin: *in vitro* and *in situ* inhibition studies. *Eur J Pharm Biopharm*, 55:241–246, 2003.
 24. Sasabe, H., Tsuji, A. and Sugiyama, Y., Carrier-mediated mechanism for the biliary excretion of the quinolone antibiotic grepafloxacin and its glucuronide in rats. *J Pharmacol Exp Ther*, 284:1033–1039, 1998.
 25. Sasabe, H., Kato, Y., Suzuki, T., Itose, M., Miyamoto, G. and Sugiyama, Y., Differential involvement of Multidrug resistance-associated protein 1 and P-glycoprotein in tissue distribution and excretion of grepafloxacin in mice. *J Pharmacol Exp Ther*, 310: 648–655, 2004.
 26. Schinkel, A.H., Smit, J J.M., van Tellingen, O., Beijnen, J.H., Wagenaar, E., van Deemter, L., Mol, C.A.A.M., van der Valk, M.A., Robanus-Maandag, E.C., te Riele, H.P.J., Berns, A.J.M. and Borst, P., Disruption of the mouse *mdr 1a* P-glycoprotein gene leads to a deficiency in the blood-brain barrier

- and to increased sensitivity to drugs. *Cell*, 77:491–502, 1994.
27. Schinkel, A.H., P-Glycoprotein, a gatekeeper in the blood–brain barrier. *Adv Drug Deliv Rev*, 36:179–194, 1999.
 28. Schrickx, J. and Fink-Gremmels, J., P-glycoprotein-mediated transport of oxytetracycline in the Caco-2 cell model. *J Vet Pharmacol Ther*, 30:25–31, 2007.
 29. Schrickx, J.A. and Fink-Gremmels, J., Danofloxacin-mesylate is a substrate for ATP-dependent efflux transporters. *Br J Pharmacol*, 150:463–469, 2007.
 30. Schuetz, E.G., Yasuda, K., Arimori, K. and Schuetz, J.D., Human MDR1 and mouse *mdr1a* P-glycoprotein alter the cellular retention and disposition of erythromycin, but not of retinoic acid or benzo(a)pyrene. *Arch Biochem Biophys*, 350:340–347, 1998.
 31. Schwarz, U.I., Gramatte, T., Krappweis, J., Oertel, R. and Kirch, W., P-glycoprotein inhibitor erythromycin increases oral bioavailability of talinolol in humans. *Int J Clin Pharmacol Ther*, 38:161–167, 2000.
 32. Seral, C., Carryn, S., Tulkens, P.M. and Van Bambeke, F., Influence of P-glycoprotein and MRP efflux pump inhibitors on the intracellular activity of azithromycin and ciprofloxacin in macrophages infected by *Listeria monocytogenes* or *Staphylococcus aureus*. *J Antimicrob Chemother*, 51:1167–1173, 2003.
 33. Sikri, V., Pal, D., Jain, R., Kalyani, D. and Mitra, A.K., Cotransport of macrolide and fluoroquinolones, a beneficial interaction reversing P-glycoprotein efflux. *Am J Ther*, 11:433–442, 2004.
 34. Sugiyama, Y., Kusuhara, H. and Suzuki, H., Kinetic and biochemical analysis of carrier-mediated efflux of drugs through the blood–brain and blood–cerebrospinal fluid barriers: importance in the drug delivery to the brain. *J Control Release*, 62:179–186, 1999.
 35. Suzuki, H., Terasaki, T. and Sugiyama, Y., Role of efflux transport across the blood–brain barrier and blood–cerebrospinal fluid on the disposition of xenobiotics in the central nervous system. *Adv Drug Deliv Rev*, 25:257–285, 1997.
 36. Takano, M., Hasegawa, R., Fukuda, T., Yumoto, R., Nagai, J. and Murakami, T., Interaction with P-glycoprotein and transport of erythromycin, midazolam and ketoconazole in Caco-2 cells. *Eur J Pharm*, 358:289–294, 1998.
 37. Tamai, I. and Tsuji, A., Transporter-mediated permeation of drugs across the blood–brain barrier. *J Pharm Sci*, 89:1371–1388, 2000.
 38. Van Bambeke, F., Michot, J.-M. and Tulkens, P.M., Antibiotic efflux pumps in eukaryotic cells: occurrence and impact on antibiotic cellular pharmacokinetics, pharmacodynamics and toxicodynamics. *J Antimicrob Chemother*, 51:1067–1077, 2003.
 39. Wakasugi, H., Yano, I., Ito, T., Hashida, T., Futami, T., Nohara, R., Sasayama, S. and Inui, K., Effect of clarithromycin on renal excretion of digoxin: interaction with P-glycoprotein. *Clin. Pharmacol. Ther*, 64:123–128, 1998.
 40. Wang, H., Liao, Z.-X., Chen, M. and Hu, X.-L., Effects of hepatic fibrosis on ofloxacin pharmacokinetics in rats. *Pharmacol Res*, 53:28–34, 2006.
 41. Weiss, M. and Kang, W. P-Glycoprotein inhibitors enhance saturable uptake of idarubicin in rat heart: Pharmacokinetic/pharmacodynamic modeling. *J Pharmacol Exp Ther*, 300: 688–694, 2002.
 42. Wong, H.L., Bendayan, R., Rauth, A.M., Xue, H.Y., Babakhanian, K. and Wu, X.Y., A mechanistic study of enhanced doxorubicin uptake and retention in multidrug resistant breast cancer cells using a polymer-lipid hybrid nanoparticle system. *J Pharmacol Exp Ther*, 317:1372–1381, 2006.
 43. Haritova, A., Schrickx, J., Lashev, L. and Fink-Gremmels, J., ABC efflux transporters – the 3rd dimension in kinetics not only of fluoroquinolones. *Bulg J Vet Med*, 9:223–242, 2006.