

Protocols for Polymersome

Content

- 1- Preparation of PMOXA-PDMS-PMOXA polymersome
- 2- Preparation of (PEG-b-P(DEAEMA-stat-BMA)) polymersome
- 3- Release profile for Doxorubicin for pH sensitive polymer
- 4- Incorporation and encapsulation of biological entities
- 5- Preparation of smart Nano-container
- 6- Characterisation

1- Preparation of PMOXA-PDMS-PMOXA polymersome

We followed as described in Corinne Nardin et.al [1], in brief, following steps were carried out,

- a. 5ml pear shaped flask is washed with pure ethanol and dried at 120°C for 1hr
- b. 1 mg polymer was dissolved in 1mL ethanol
- c. Solution was evaporated at 40°C (water bath temperature), 40 rpm (rotation speed at an angle as high as possible, ~ 60°-70°) and a pressure of 150 mbar in rotary evaporator
- d. After complete evaporation of ethanol, pressure was reduced using rotary pump vacuum ($\sim 10^{-2}$ mbar) for 30 min.
- e. Later 1mL PBS (pH 7.4) was added and stirred overnight at 300rpm
- f. In order to remove residual lamellar material and to define particle size less than 200nm, samples were extruded using ultra-filtration equipment (Avanti Polar mini Extruder) with Nucleopore membrane (0.2 μ m pore size)

2- Preparation of (PEG-b-P(DEAEMA-stat-BMA)) polymersome

We followed as described in Mohamed A. Yassin, et.al [2], in brief, following steps were carried out,

- a. Millipore water was used for solution preparation
- b. 1mg Polymer was dissolved in HCl (pH 2) with the help of magnetic stirrer at 700rpm
- c. Later the pH of solution was reduced from 2 to 8, by adding NaOH drop wise at very slow rate. The pH of the solution should be checked very carefully.

3- Release profile assay

It was performed using cellulose dialysis membrane of MWCO 5000Da and 300KDa for biological entities less than 500Da and Enzymes respectively

- a. The beaker was filled with dialysis solution (Milli-Q wáter or Buffer solution)
- b. Dialysis tube was cut as per the volume of sample and dialysis tube width.
- c. The tube was washed in Milli-Q wáter and pre-incubated in Milli-Q wáter for

20mins.

- d. After pre-incubation, the tube was closed on 1 end with magnetic clamp such that atleast 10mm tubing overlaps.
- e. Then through the other end, sample was filled inside the tube and air was removed carefully before closing the tube with clamp as before.
- f. Dialysis tube was placed in prepared beaker and stirred at low speed (300rpm)
- g. Dialysis solution was replaced in beaker after every 2 hour
- h. After each hour, sample's were analysed using Beckman Coulter – UV 800 Spectrophotometer for Doxorubicin, enzyme, Tamra dye, fluorescein dye at 480nm, 280nm, 555nm and 490nm respectively
- i. After dialysis, the sample was removed carefully after drying the tube with kim wipe.

4- Incorporation and encapsulation of biological entities

Incorporation of DNA channel (15nM conc.) and Encapsulation of enzymes (20% of polymer conc.) was carried out by adding during each polymersome preparation in following ways,

PMOXA-PDMS-PMOXA: DNA channel or Enzyme was added to PBS buffer in last step along with 5µl MgCl₂ solution (1 M)

(PEG-b-P(DEAEMA-stat-BMA)): Polymersome solution pH was brought to 5 and DNA channel (15nM conc.) or Enzyme was added along with 5µl MgCl₂ solution (1 M). Further, solution pH was reduced further to 8.

5- Preparation of smart Nano-container

Each polymersome was prepared along with DNA channel (15nM Conc.) and Enzyme as follows,

PMOXA-PDMS-PMOXA: DNA channel followed by Enzyme was added to PBS buffer in last step along with 5µl MgCl₂ solution (1 M)

(PEG-b-P(DEAEMA-stat-BMA)): Polymersome solution pH was brought to 5 and Enzyme was added and after few minute, DNA channel (15nM conc.) was added along with 5µl MgCl₂ solution (1 M). Further, solution pH was reduced further to 8.

6- Characterisation

The following instrument were used to characterize the polymersome,

- a. Cryo-TEM (Transmission Electron Microscope)
 - i) For frozen hydrated specimen preparation, 2µl of sample was deposited on a holey carbon support film attached to a copper grid.

- ii) The liquid film was thinned by blotting for 0.5 sec and frozen rapidly in liquid ethane.
 - iii) After rapid freezing, the sample was loaded to a Gatan cryo transfer holder keeping temperature below -130°C in order to prevent ice crystallization.
 - iv) TEM inspection was done in ZEISS LIBRA 120, energy filtered images were recorded under low dose electron irradiation conditions.
 - v) Samples were kept below -130°C during the inspection.
- b. Cryo-SEM (Scanning Electron Microscope)
- i) Samples were rapidly frozen in liquified ethane according to standard procedure.
 - ii) Frozen samples were fractured at approximately -130°C in a cryo preparation chamber and transferred to the cooling stage inside a ZEISS Ultra (.....) after coating with 3nm Carbon layer.
- c. DLS (Dynamic Light Scattering)
- Particle size analysis of aqueous solutions of polymerosomes and gold nanoparticles was performed by dynamic light scattering using a Malvern Zetasizer Nano. Data analysis is based on non negative least squares (NNLS) method [3].

Reference:

1. Corinne Nardin, Thomas Hirt, Jorg Leukel, and Wolfgang Meier, Polymerized ABA Triblock Copolymer Vesicles, *Langmuir* 2000, 16, 1035-104
2. Mohamed A. Yassin, Dietmar Appelhans, Rafael G. Mendes, Mark H. Rummeli, and Brigitte Voit, *Chem. Eur. J.* 2012, 18, 12227 – 12231
3. Robert Finsy, Nicolas de Jaeger, Rick Sneyers and Eric Gelade *Part. Part. Sys. Charact.* 1992, 9, 125