

Got Milk?

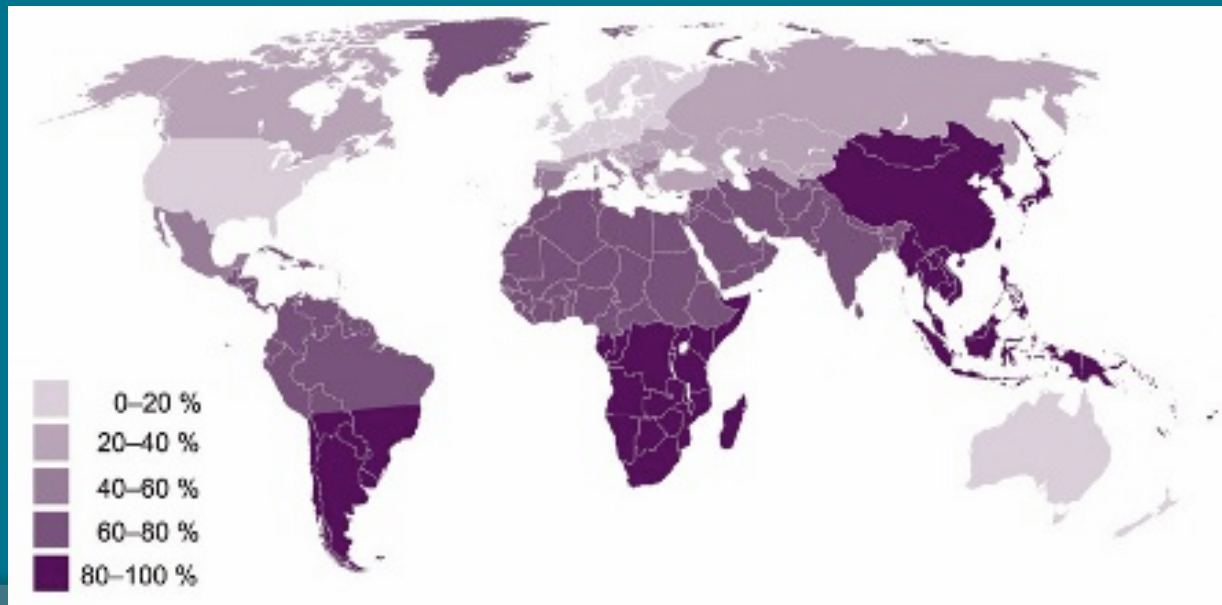
Team Übermensch

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Mentors: Derek Ju and Alie Doolittle

Lactose Intolerance

- Lack the enzyme lactase due to environmental or genetic factors
- Affects 70-75% of the worldwide population
- Varies with ethnicity

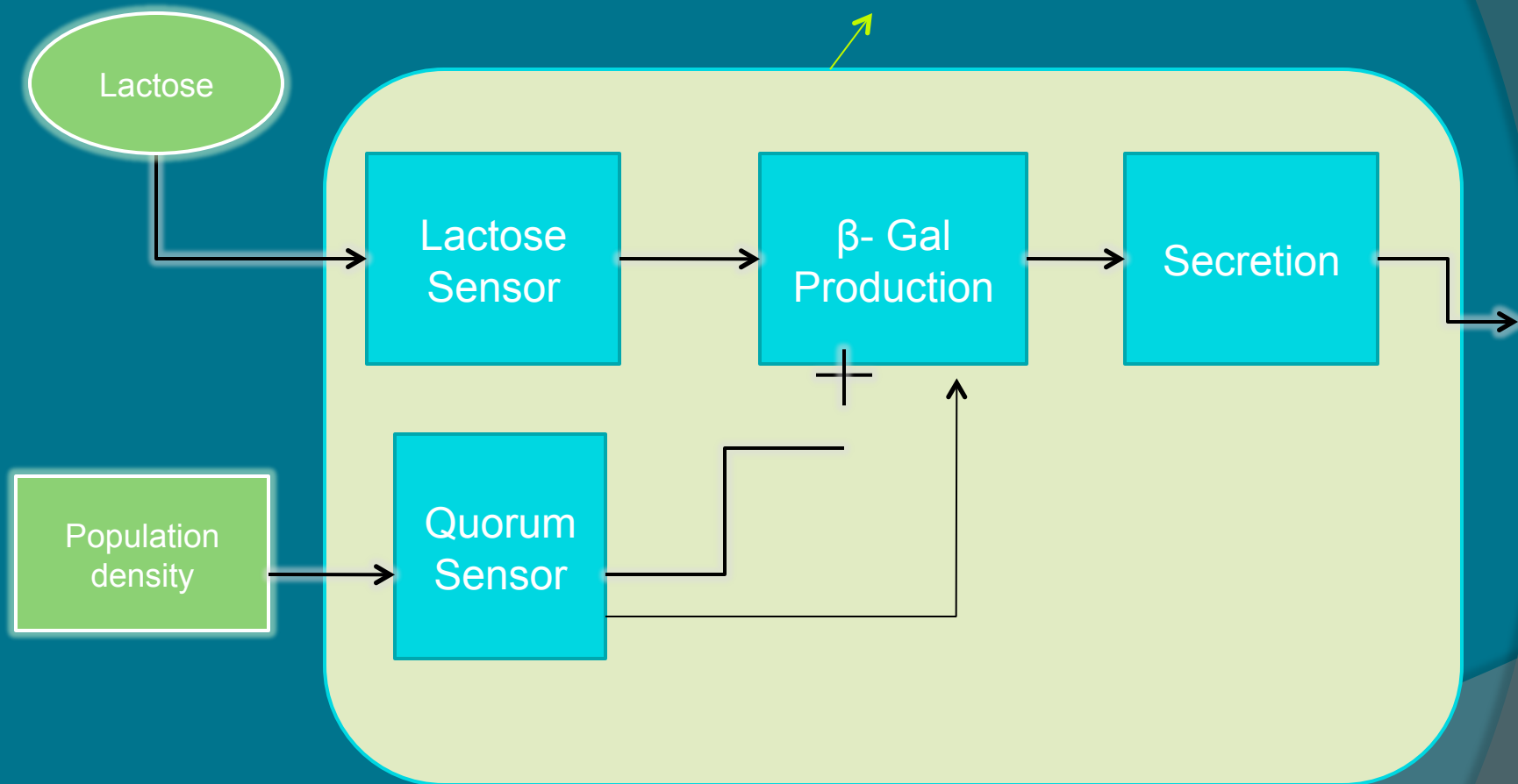


Overall System

- Bacterial production of β -Galactosidase when lactose is present in human digestive system
- Maintenance of bacterial population in the GI tract at functional numbers

High Level System Overview

Bacterial Cell



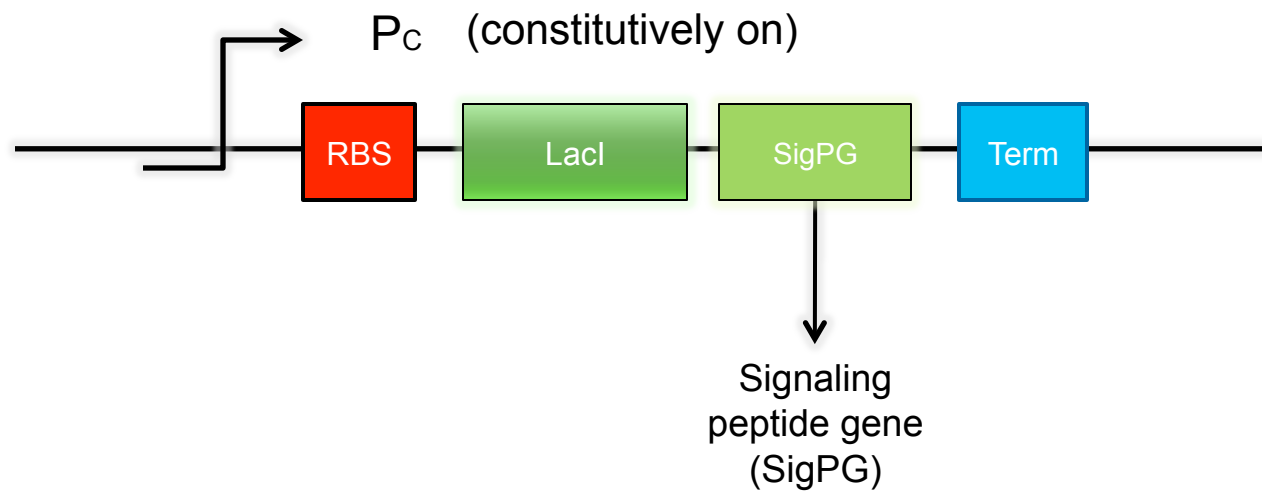
Inputs and Outputs

	Lactose On	Lactose Off
High Population	Produce β -Gal	Normal Replication
Low Population	Stop producing β -Gal, Normal Replication	Normal Replication

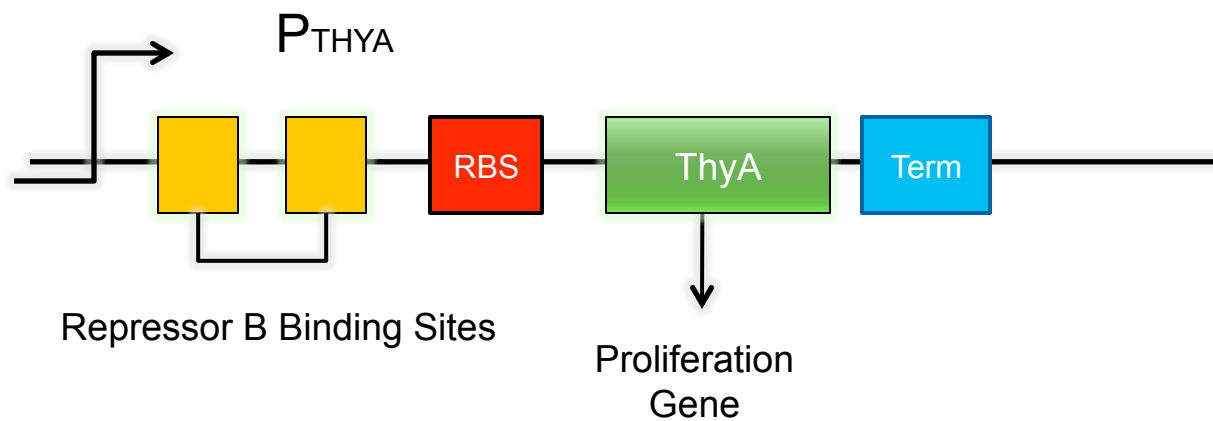


DEVICES

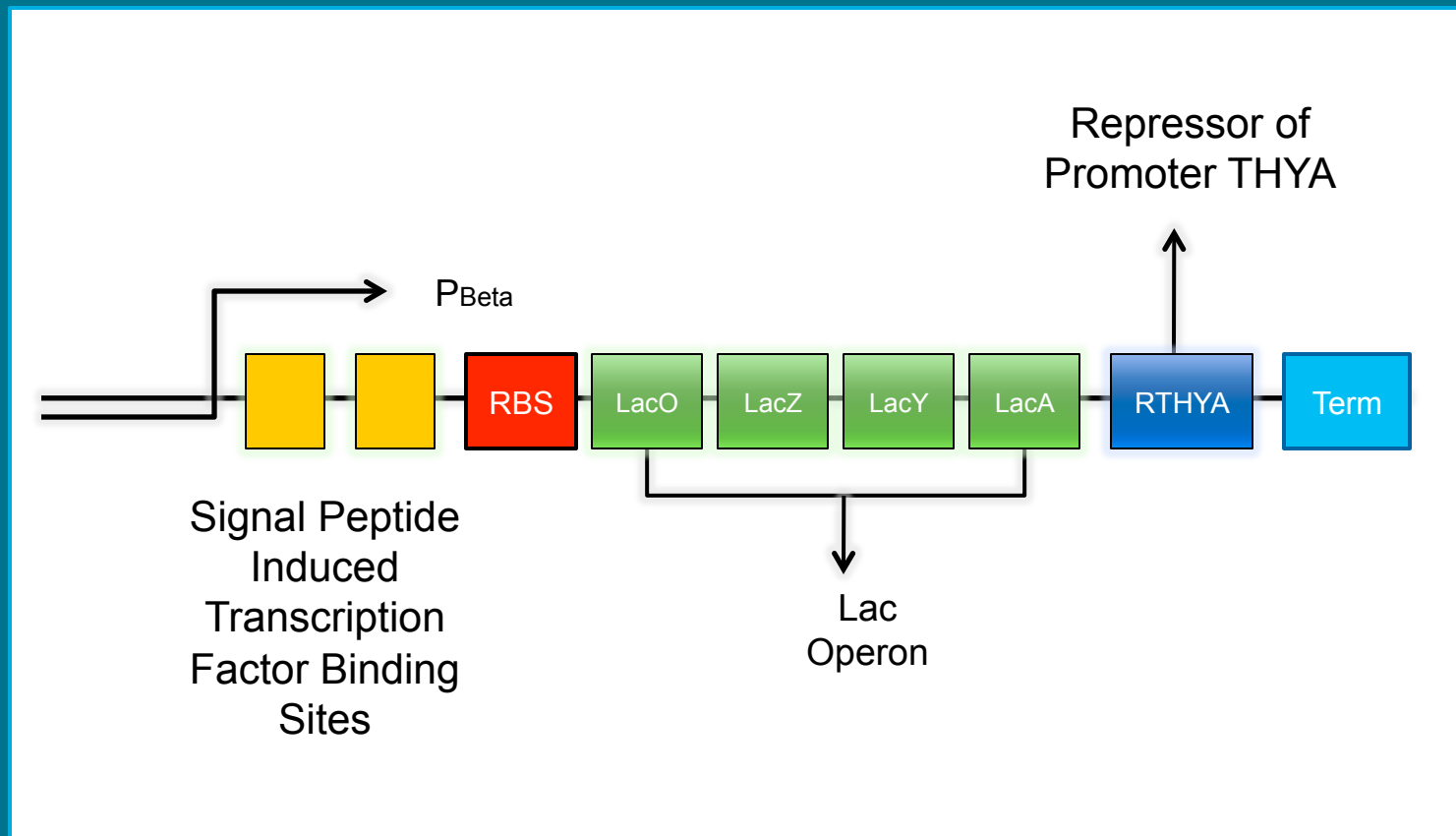
Lactose Sensor



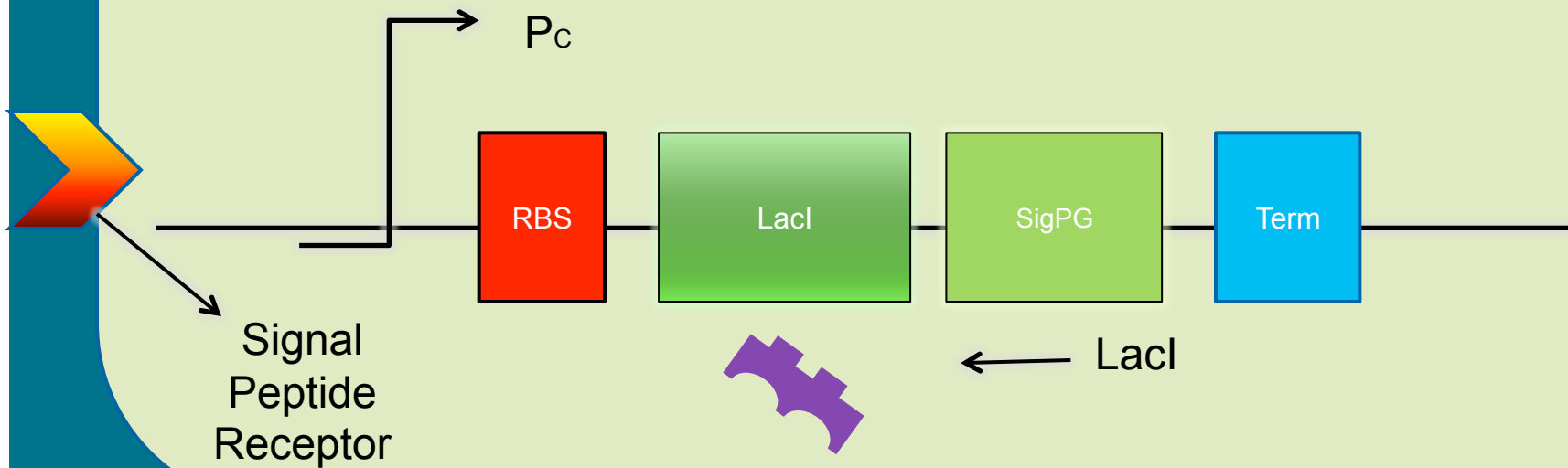
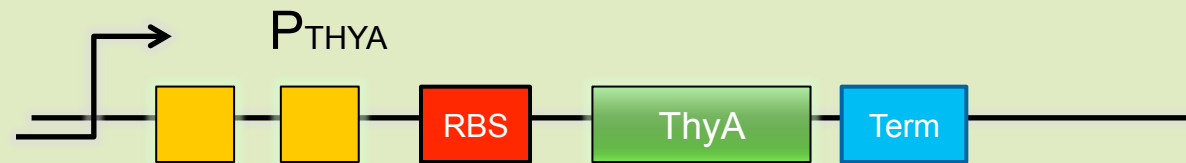
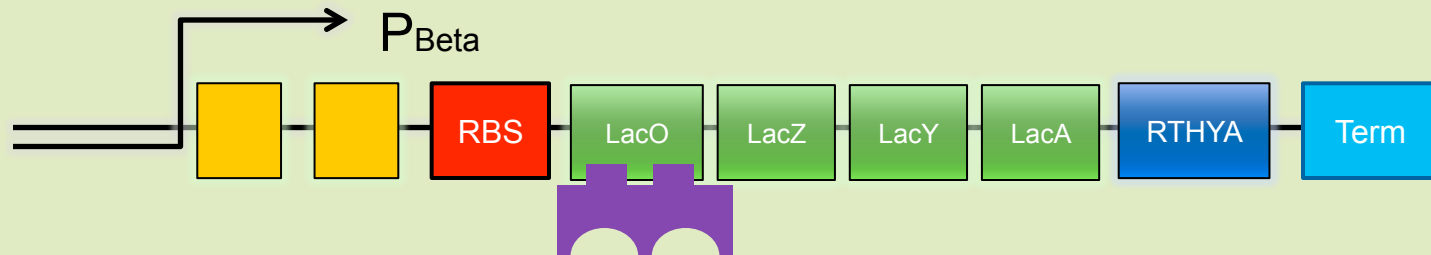
Quorum Sensor



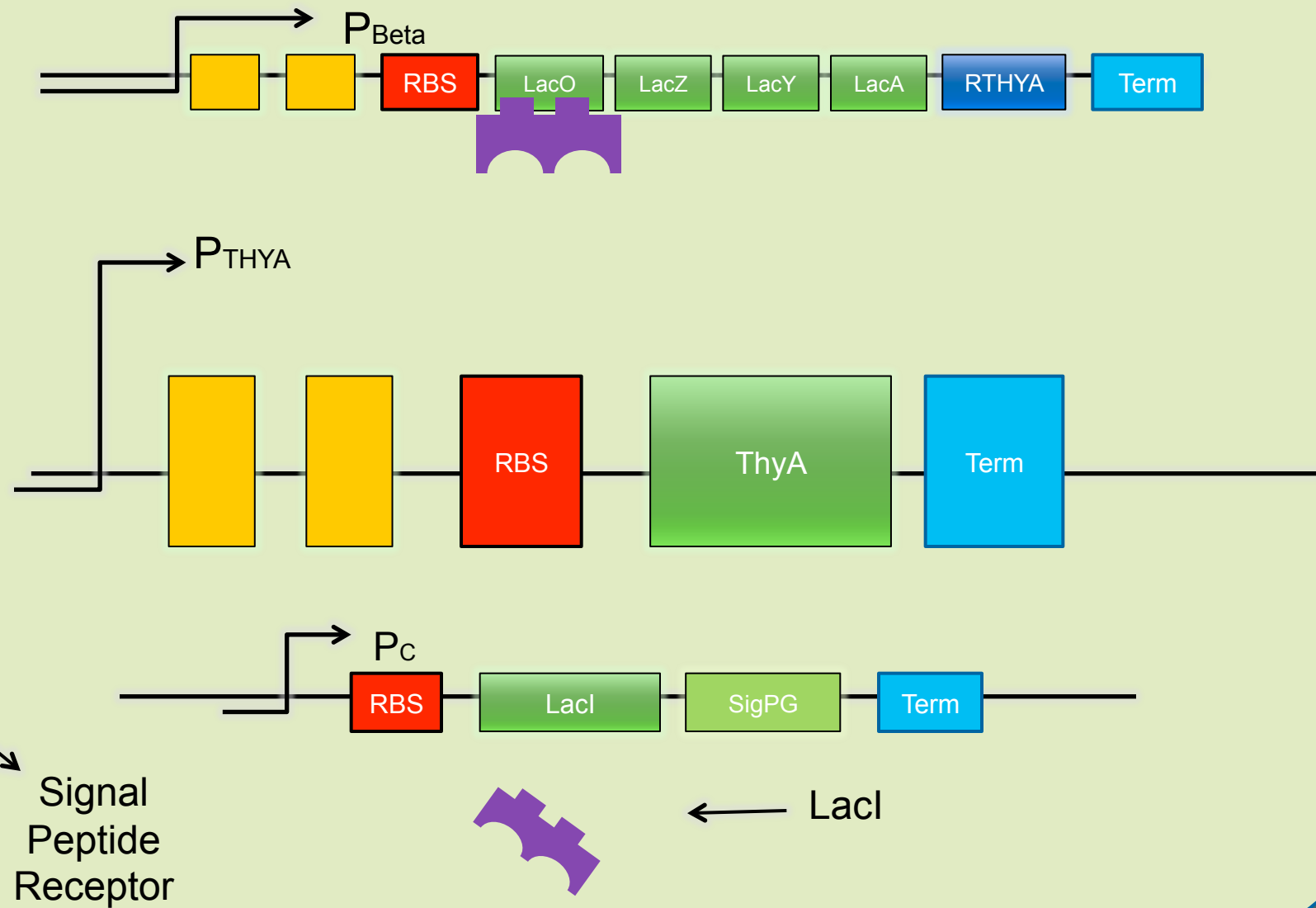
β -Galactosidase Production



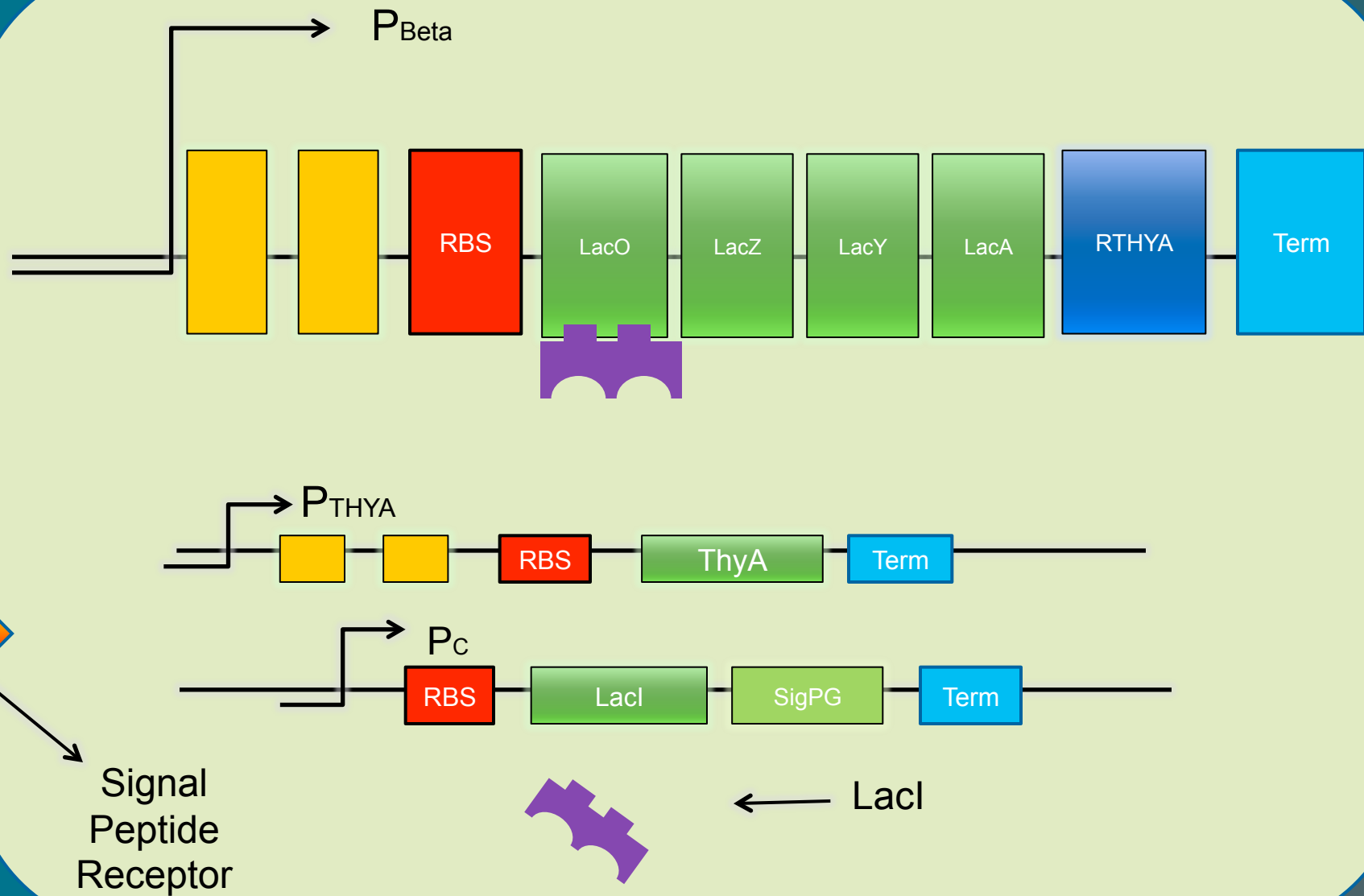
System



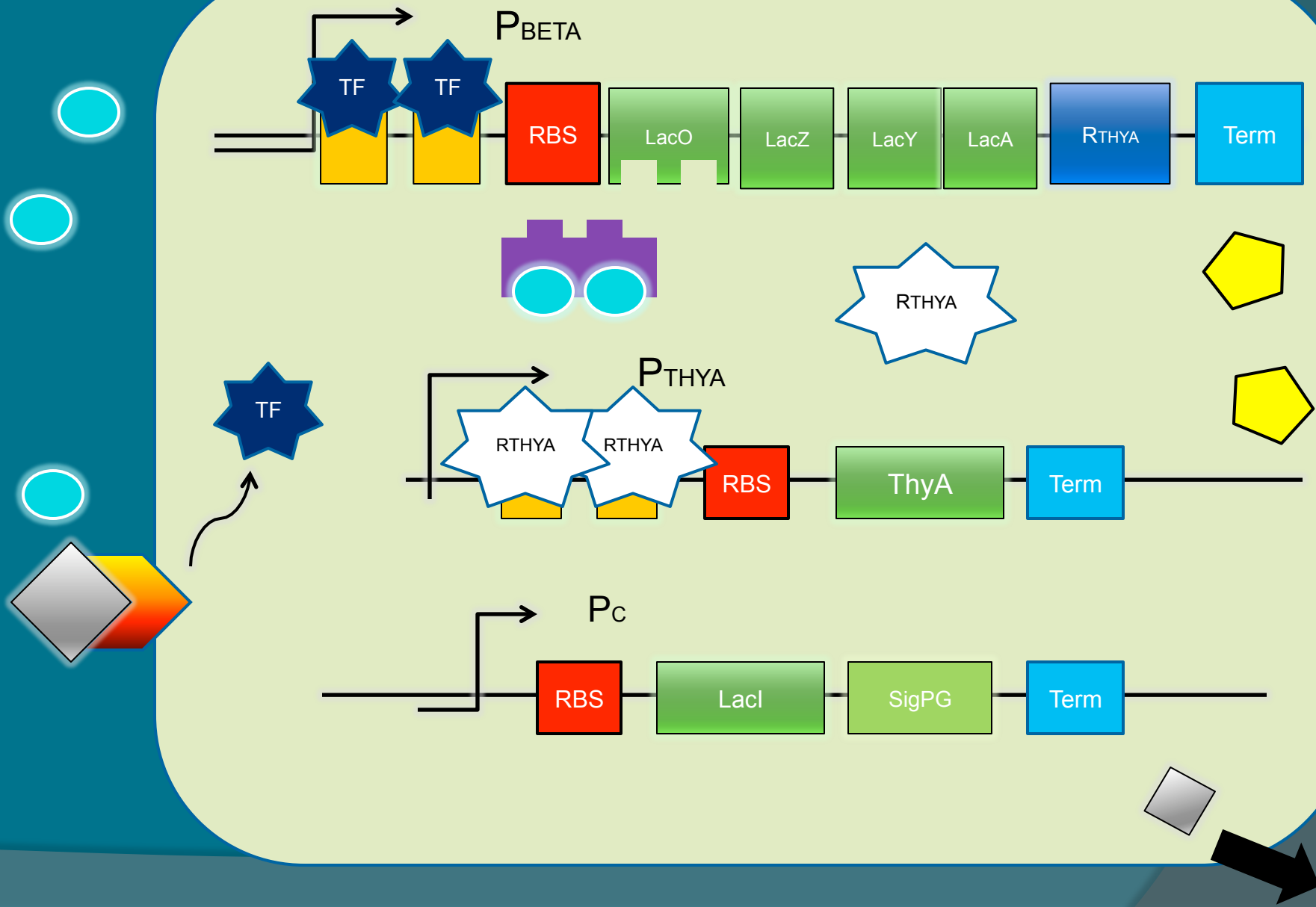
System



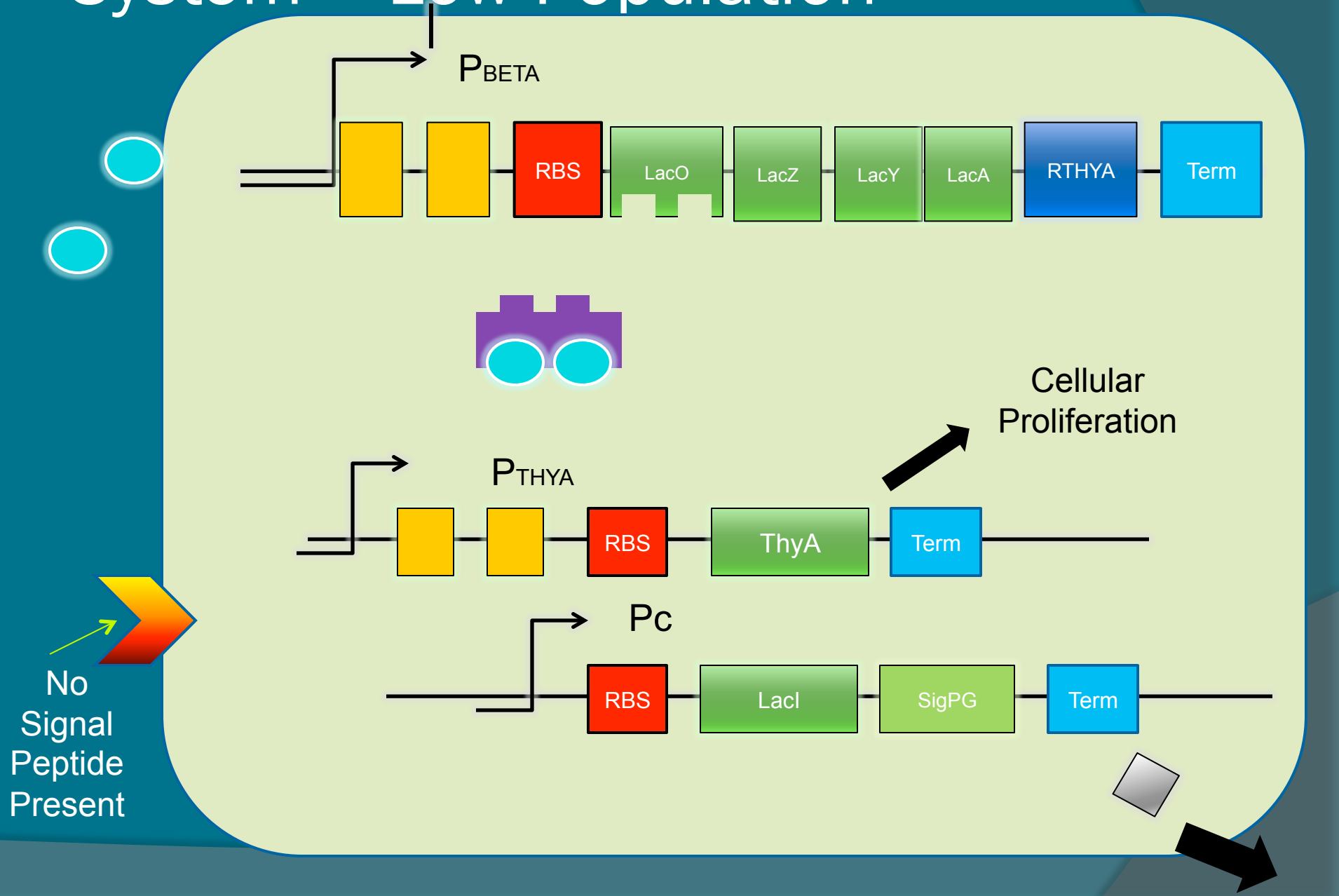
System



System - High Population

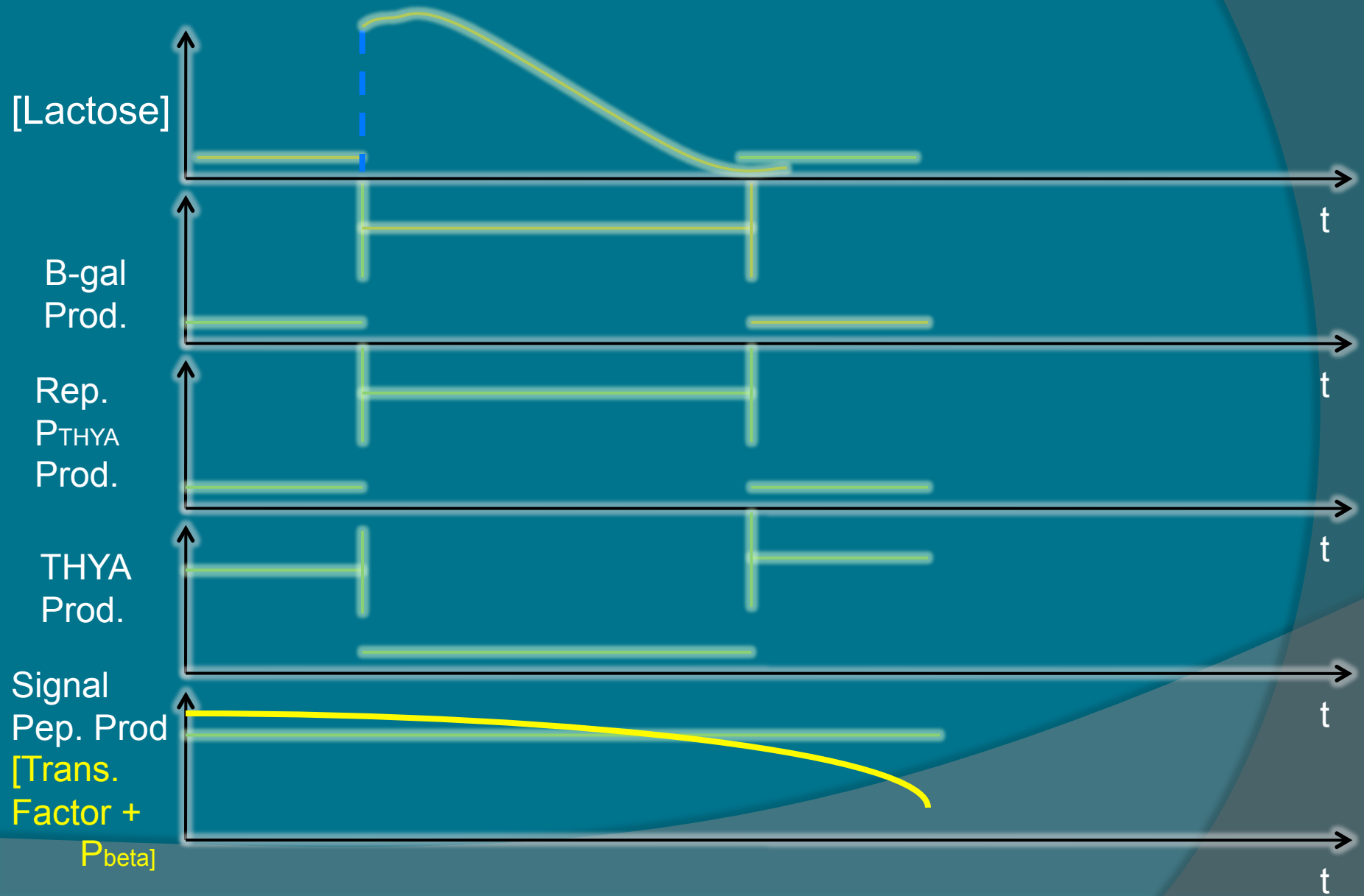


System – Low Population



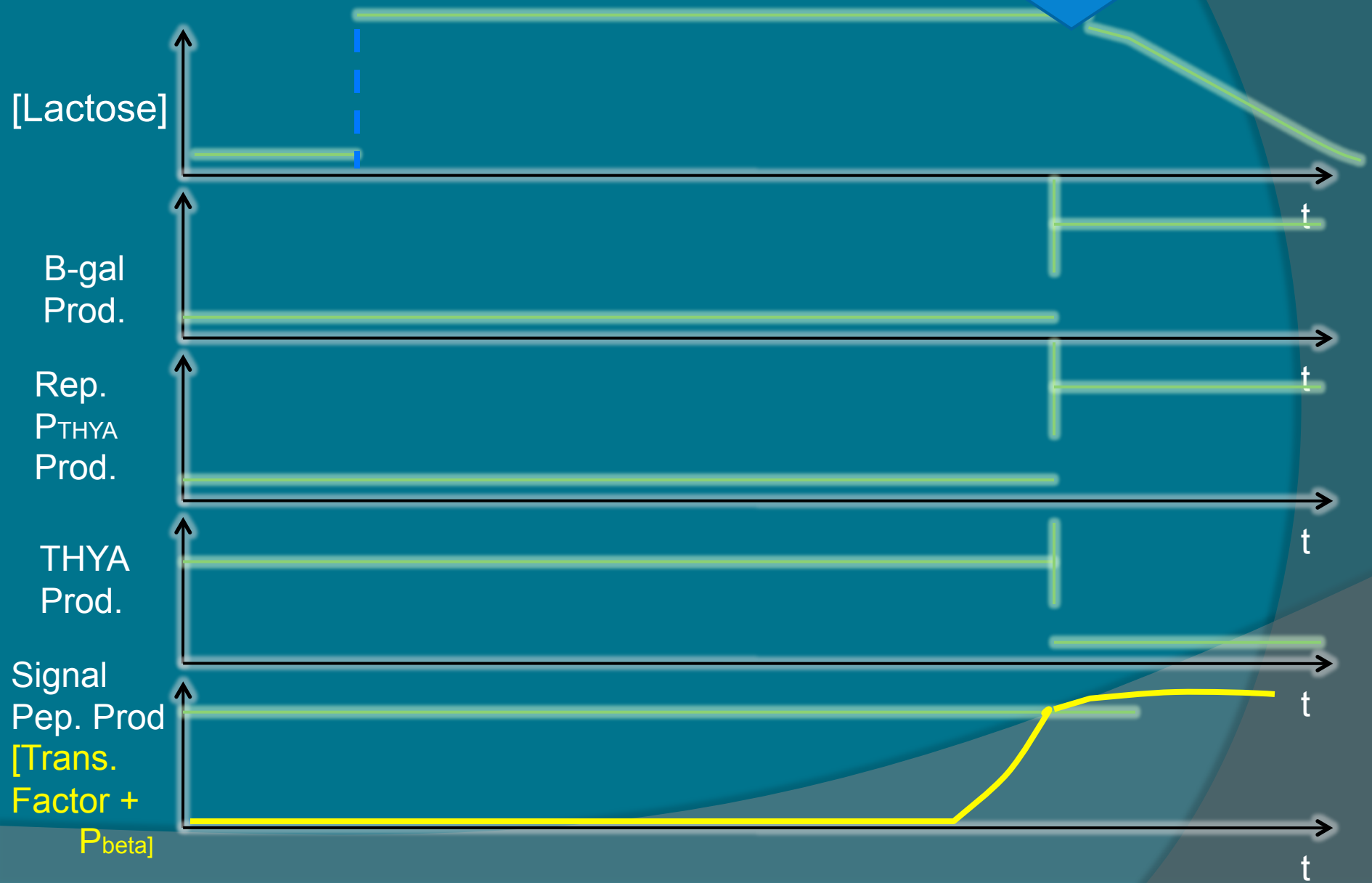
TIME DIAGRAM

Time Diagram: High Population



Time Diagram: Low pop

High
Level
Pop.



TESTING/DEBUGGING

Strategy for Testing: general cloning

- ⦿ Restriction Enzyme Digest and Gel Electrophoresis
- ⦿ Sequence plasmid after each insertion to check transformation of DNA
 - Check for mutations
 - Knockout- Auxotrophs
 - Antibiotic resistance

Strategies for Testing: β -Gal Production/Function

● Test for Devices

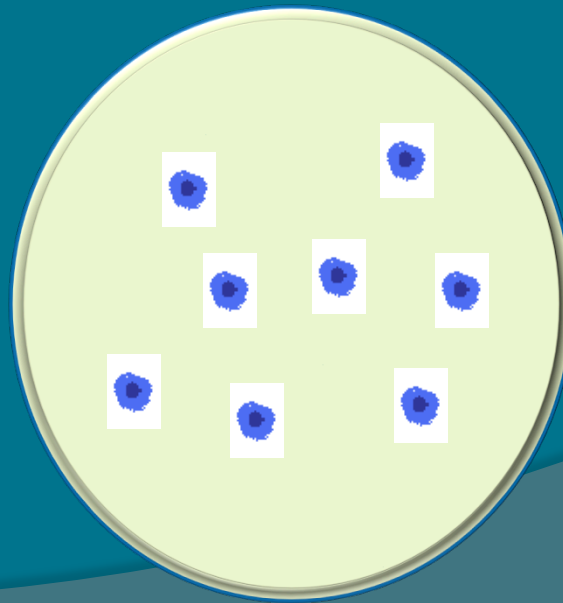
- GFP-tag a component in each device
- Fluorescence indicator
- Promoter activity



-Protein activity: isolate proteins and test activity *in vitro*

Strategies for Testing: Secretion

- Grow on X gal culture
- The agar plate around the colonies with correct gene should turn blue



Strategies for Testing: Quorum Sensing

- Signal Peptide production?
 - Tag it with GFP
- Signal Peptide interacts with receptor → transcription factor?
 - Insert Signal peptide, tag transcription factor with GFP: does it produce the transcription factor?
- Transcription factor + P_{beta} ?: should produce β -Gal

Equations

- $\text{RNAPol} + \text{Pc} \rightarrow \text{LacI mRNA} + \text{sigPG mRNA}$
- $\text{LacI mRNA} + \text{ribosome} \rightarrow \text{LacI}$
- $\text{LacI mRNA} \rightarrow \text{decay}$
- $\text{SigPG mRNA} + \text{ribosome} \rightarrow \text{SigPG}$
- $\text{SigPG mRNA} \rightarrow \text{decay}$
- $\text{LacI} + \text{LacO} \rightarrow \text{LacILacO}$
- $\text{LacILacO} + \text{Allolactose} \rightarrow \text{LacIAllolactose} + \text{LacO}$
- $\text{SigPG} + \text{receptor} \rightarrow \text{SigPGreceptor} + \text{TF}$
- $\text{TF} + \text{P}_{\text{beta}} \rightarrow \text{TFP}_{\text{beta}}$
- $\text{TF} + \text{TFP}_{\text{beta}} \rightarrow 2\text{TFP}_{\text{beta}}$
- $2\text{TFP}_{\text{beta}} + \text{RNAPol} \rightarrow \beta\text{-Gal mRNA} + \text{R}_{\text{THYA}} \text{ mRNA}$
- $\beta\text{-Gal mRNA} + \text{RNAPol} \rightarrow \beta\text{-Gal}$

Equations

- $\beta\text{-Gal mRNA} \rightarrow \text{decay}$
- $R_{\text{THYA}} \text{ mRNA} + \text{ribosome} \rightarrow R_{\text{THYA}}$
- $R_{\text{THYA}} \text{ mRNA} \rightarrow \text{decay}$
- $R_{\text{THYA}} + P_{\text{THYA}} \rightarrow RP_{\text{THYA}}$
- $R_{\text{THYA}} + RP_{\text{THYA}} \rightarrow 2RP_{\text{THYA}}$
- $2RP_{\text{THYA}} \rightarrow RP_{\text{THYA}} + R_{\text{THYA}}$
- $RP_{\text{THYA}} \rightarrow R_{\text{THYA}} + P_{\text{THYA}}$
- $P_{\text{THYA}} + \text{RNApol} \rightarrow \text{ThyA}$
- $2\text{TFP}_{\text{beta}} \rightarrow \text{TF} + \text{TFP}_{\text{beta}}$
- $\text{TFP}_{\text{beta}} \rightarrow \text{TF} + P_{\text{beta}}$
- $\text{SigPGreceptor} \rightarrow \text{SigPG} + \text{receptor}$
- $\text{TF} \rightarrow \text{decay}$
- $R_{\text{THYA}} \rightarrow \text{decay}$
- $\text{LacI} \rightarrow \text{decay}$

Cost – Biological Parts

- Most needed proteins are relatively common, most of our peptide parts will be found in the registry of biological parts.
 - 3 promoters, ~45 bp each, \$.40/bp
 - 5 pairs of primers, 20-40 bp each, \$.40/bp
 - DNA synthesis, \$400
 - \$255 for a strain of *Lactobacillus acidophilus*
-

Grand Total: ~\$1,000*

*Unless we learn that we need proteins that are not in the registry.

A background image showing a stream of white milk being poured from a white container into a clear glass. The milk is captured mid-pour, creating a dynamic sense of movement. The glass is partially filled with milk, and the background is a soft, out-of-focus light beige.

Benefits

People will be able to ingest lactose containing products without regular ingestion of lactase pills or lactose-free products!

Unanswered Questions

- 1) How do you get the synthetic genes into the genome?
- 2) How do you isolate the appropriate signal peptide, transcription factor and promoter that it would interact with?
- 3) Can we rely on the natural bacterial secretory pathway?
- 4) Can we assume that the chassis will not over-proliferate due to the natural competition in lower GI?
- 5) Kill switch: should we have one in order to eliminate those with too many mutations on the inserted genes?

⦿ Buildable?

- Yes!

⦿ Cost?

- Reasonable!

⦿ Safe/Secure?

- Yes! (BL1 microorganism)

Go or No Go?



GO!

References

- ◉ http://thelazygfcchef.files.wordpress.com/2010/01/1056910_51209851.jpg
- ◉ <http://raceforareason2009.files.wordpress.com/2009/04/340x.jpg>
- ◉ www.atcc.org
- ◉ <http://mic.sgmjournals.org/cgi/content/abstract/153/12/3939>
- ◉ <http://aem.asm.org/cgi/content/full/73/13/4259?maxtoshow=&hits=10&RESULTFORMAT=&fulltext=harveyi&searchid=1&FIRSTINDEX=40&resourcetype=HWFID>
- ◉ <http://nccam.nih.gov/health/probiotics/>