

# Module Overview

Day	Lecture	Lab
1	Introduction	DNA library synthesis (PCR)
2	SELEX I: Building a Library	DNA library purification (agarose gel electrophoresis)
3	SELEX II: Selecting RNA with target functionality	RNA library synthesis ( <i>In vitro</i> transcription = IVT)
4	SELEX III: Technical advances & problem-solving	RNA purification and heme affinity selection
5	Characterizing aptamers	RNA to DNA by RT-PCR
6	Introduction to porphyrins: chemistry & biology	Post-selection IVT <a href="#">Journal Club 1</a>
7	Aptamer applications in biology & technology	Aptamer binding assay
8	Aptamers as therapeutics	<a href="#">Journal Club 2</a>

# SELEX I

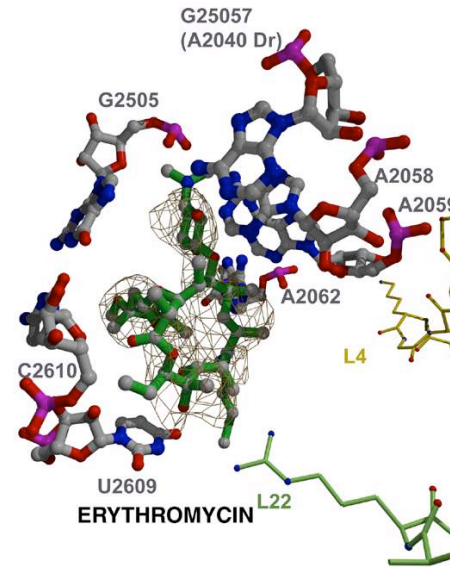
## *Building a Library*

20.109 Lecture 2

8 February, 2011

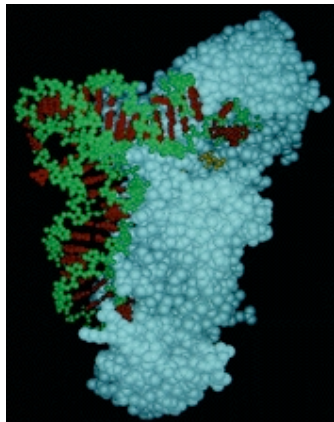
# Last time ...

## Defined RNA-small molecule interactions

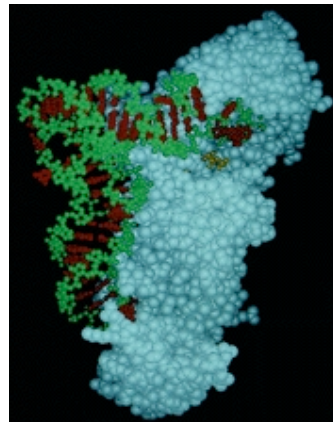


23S rRNA: erythromycin

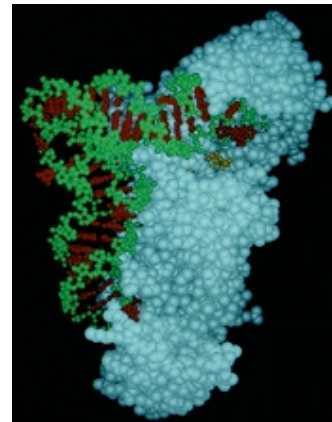
## Unique RNA-protein interactions



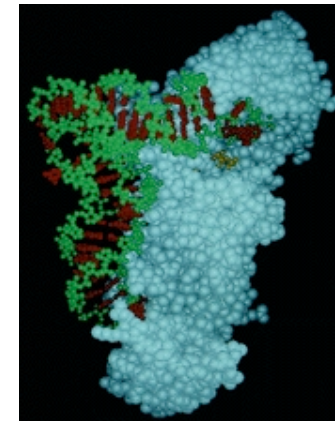
tRNA (1)  
aaRS (1)



tRNA (2)  
aaRS (2)



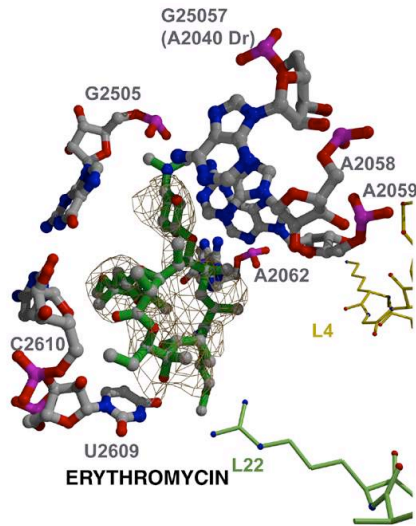
tRNA (3)  
aaRS (3)



tRNA (4)  
aaRS (4)

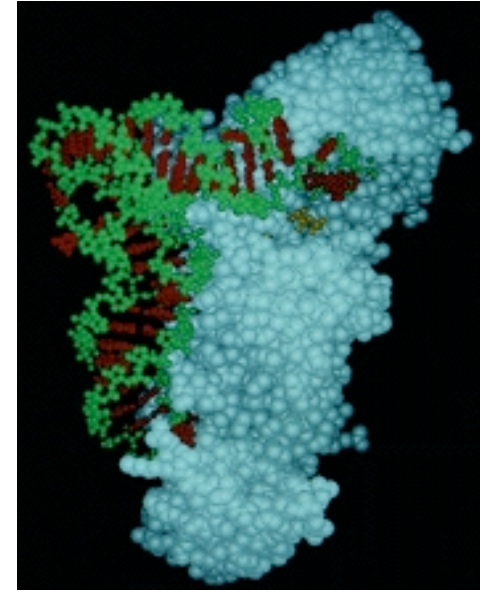
.....

# Last time ...



23S rRNA: erythromycin

- *Can we discover novel RNA molecules that interact with any target of interest?*



tRNA::aaRS

- In Nature, RNA interacts with both small molecules and proteins
- The 3D structure of the RNA permit stabilizing atomic contacts to be made
- Subtle differences in RNA 3D structure can lead to distinct binding partner interactions

# Today's Objectives

- Better conceptualize the **SELEX** process for selecting RNA **aptamers** with desired binding affinity
- Understand some basic principles influencing RNA library design
  - Appreciate how practical issues shape library architecture
  - Understand the concept of library diversity
  - Appreciate the limitations in building an ideal library

# Discovering your desired RNA

1. Design-oriented approach
2. Selection-based approach

# Discovering your desired RNA

*“Design-oriented approach”*

***Decide on target function***



***Design specific RNA to meet function***

## Challenges

1. Difficult to predetermine the RNA structure required for function
2. Cannot robustly use linear RNA sequence information to completely infer:

- Structure
- Function

## Requires

1. *A priori* knowledge of target RNA structure required for function
2. Ability to predict RNA structure based on simple inputs (e.g. sequence)

# Discovering your desired RNA

## *“Selection-oriented approach”*

**Decide on target function**



**Query RNA pool**  
(Apply selection pressure)



**Isolate RNA with  
desired activity**

### **Requires:**

1. Access to a sufficiently diverse RNA pool
  - Increased probability that the desired activity is present
2. Effective strategy for eliminating “losers” and enriching for “winners”

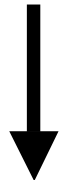


# Discovering your desired RNA

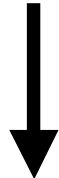
*“Selection-oriented approach”*

*Presently tenable*

Decide on target function



Query RNA pool  
(Apply selection pressure)



Isolate RNA with  
desired activity

## Advantages

1. No *a priori* knowledge of structure  $\Leftrightarrow$  function relationship required
2. Function drives emergence of a solution
  - By default, “winner” RNA has the requisite structure for function!

# Discovering RNA with novel properties

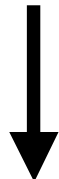
- **SELEX**

- **S**ystematic **E**volution of **L**igands by **EX**ponential enrichment
  - A selection-based strategy

Decide on target



Query RNA pool  
(Apply selection pressure)



Isolate RNA with  
desired binding activity

= **Aptamer**

– Derived from latin word  
“*aptus*” meaning “to fit”

– RNA aptamer = RNA  
derived from a large pool  
having specific binding  
affinity for a target  
molecule



**Larry Gold**  
(U. Colorado)

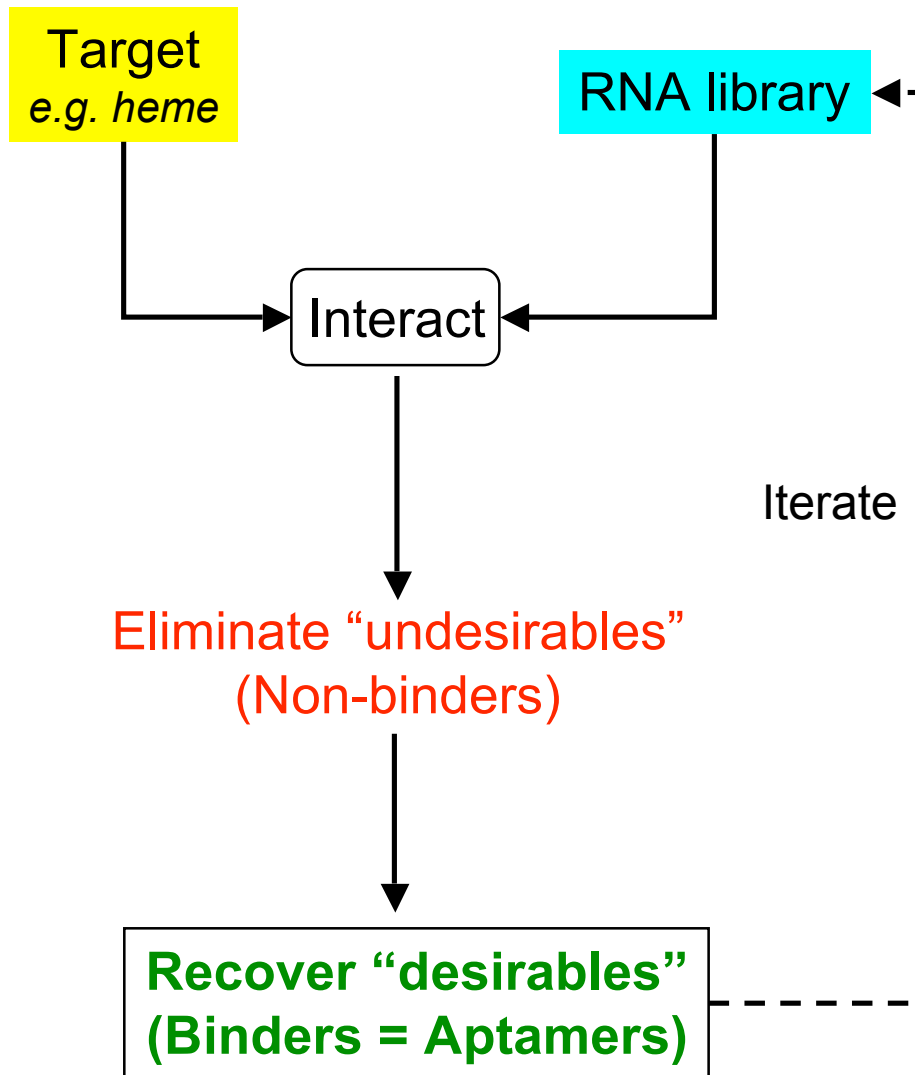


**Jack Szostak**  
(Harvard U.)

C. Tuerk and L. Gold; **Science**; 249 (4968), 505-510, 1990.

A.D. Ellington and J.W. Szostak; **Nature**; 346 (6287), 818-822, 1990.

# SELEX: The process (simply)



- **Materials:**
  - Target of interest
  - RNA library
- **Need strategies for:**
  - Exposing target to library
  - Eliminating non-binders (partitioning step)
  - Recovering binders
  - Expanding recovered pool after each round

# Conceptualizing SELEX

**Molecular targets**

*e.g. heme*

**Majors**

20

1

2

3

4

5

6

.

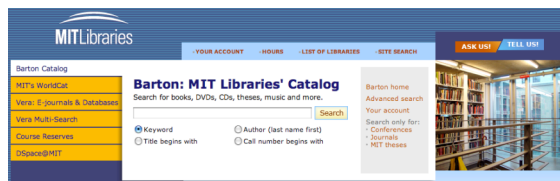
.

.

24

**SELEX**

*Strategy for efficiently  
querying your RNA library*



**Barton**

*Strategy for efficiently  
querying the MIT Collections*



History

Science

Engineering

Philosophy

**Book collection =  
RNA sequence  
collection**

**Book {x,y,z} =  
Aptamer**

Book X

Book Y

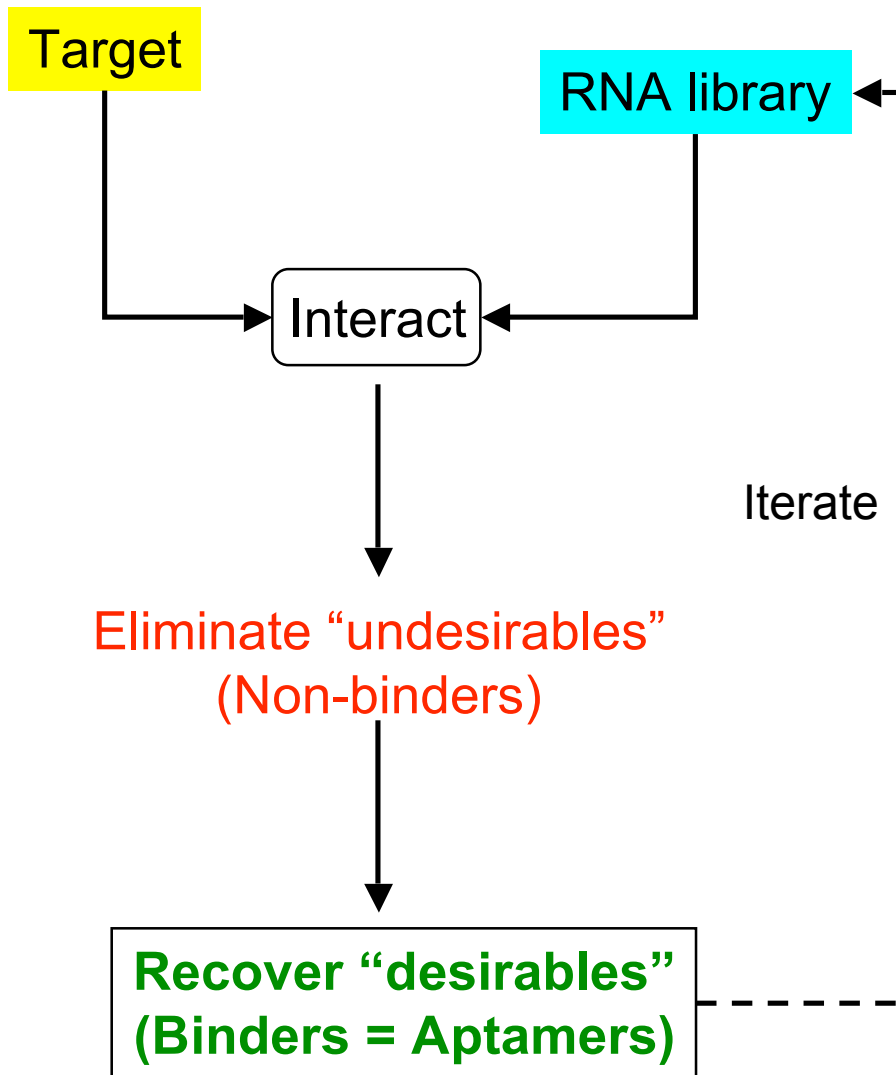
Book Z

**Student major dictates  
which books will be in  
demand**



**Populate library with "books"  
expected to be in demand by  
students**

# Target selection



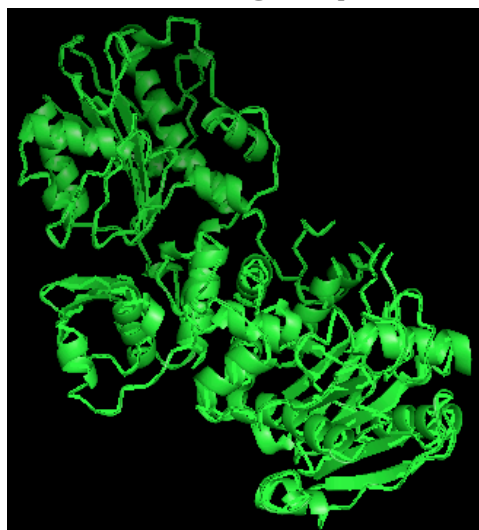
# Target selection

## Target

- The (mostly) trivial part
- Driven by investigator's interest(s)



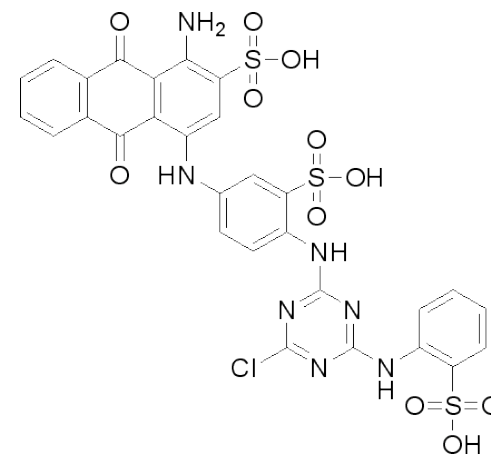
*RNA binding to protein*



*T4 DNA polymerase*  
*Residues 1-388*  
([www.rcsb.org](http://www.rcsb.org))



*RNA binding to small molecule organic dyes*

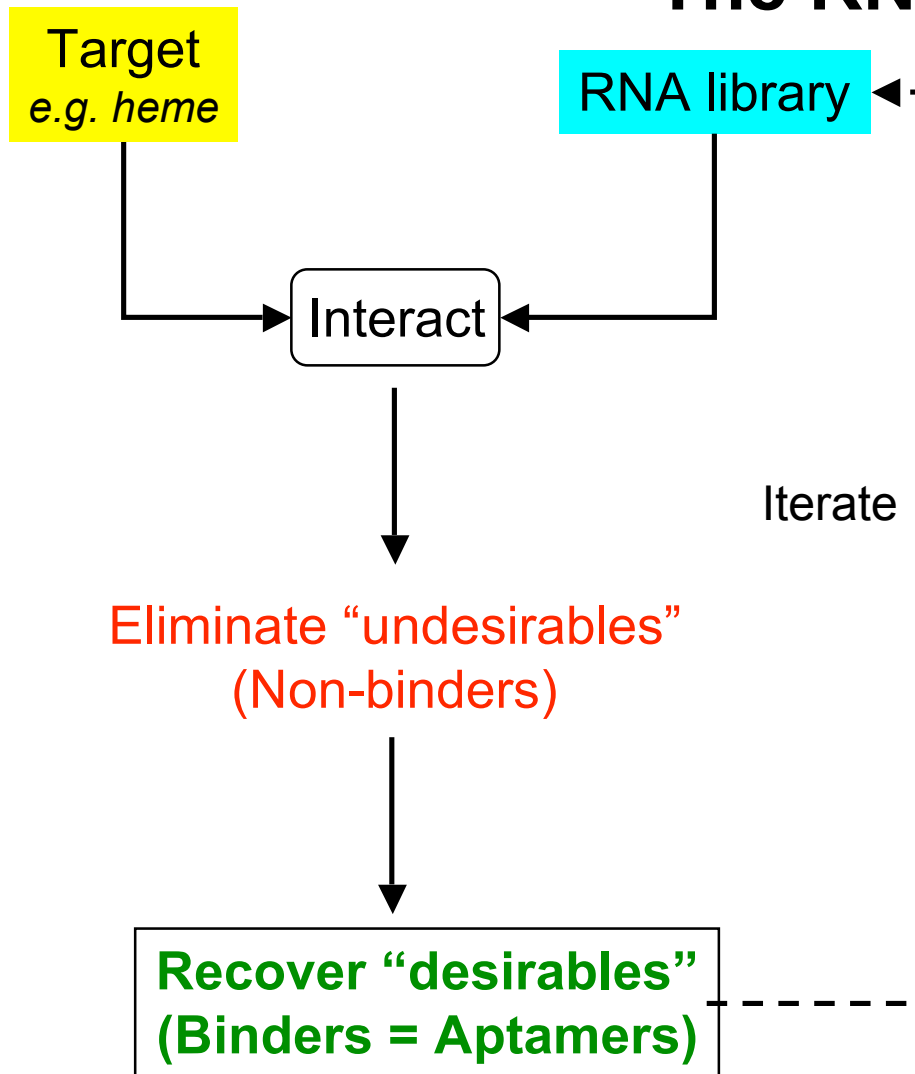


*Cibracon Blue*

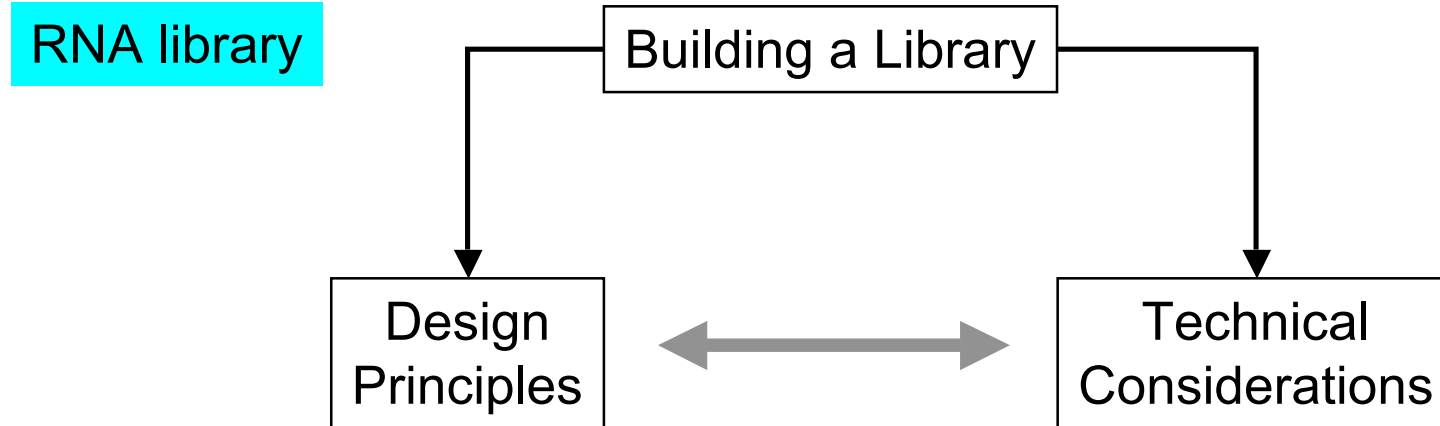
C. Tuerk and L. Gold; **Science**; 249 (4968), 505-510, 1990.

A.D. Ellington and J.W. Szostak; **Nature**; 346 (6287), 818-822, 1990.

# The RNA Library



# The RNA Library (abstracted)



- One library per target **or** *one library for all targets*
- Balance between “useful” and “useless” library members
- Maximizing “useful” collection within space constraints
- *Overall, library must be in a technical format compatible with all the steps involved in SELEX*
- Stability during storage
- Synthesizing library at reasonable costs
- Availability of efficient methods for manipulating library



# Technical considerations

- **Stability during storage**
  - DNA versus RNA?
  - DNA is more stable than RNA
    - RNA much more susceptible to hydrolysis than DNA;
      - Divalent metal catalyzed
    - RNA *highly* susceptible to ubiquitous RNases
  - *DNA is an excellent long-term form for stably storing library*

# Technical considerations

- Synthesis costs
  - DNA



[www.idtdna.com](http://www.idtdna.com)

## Custom Oligonucleotide Synthesis

Desalted custom synthesized DNA oligos are shipped lyophilized or hydrated with **Lab Ready Oligo Service**. Synthesis scales up to 1  $\mu$ mole are shipped the next business day. 5  $\mu$ mole and 10  $\mu$ mole scales are shipped within 5 business days.

Base Pricing		
Synthesis Scale	Price	
25 nmole DNA Oligo	\$0.35 USD / Base	<a href="#">Order</a>
100 nmole DNA oligo	\$0.55 USD / Base	<a href="#">Order</a>
250 nmole DNA oligo	\$0.95 USD / Base	<a href="#">Order</a>
1 $\mu$ mole DNA oligo	\$1.95 USD / Base	<a href="#">Order</a>
5 $\mu$ mole DNA oligo	\$9.50 USD / Base	<a href="#">Order</a>
10 $\mu$ mole DNA oligo	\$17.50 USD / Base	<a href="#">Order</a>

- DNA oligo 100 bases long
- 1  $\mu$ mol scale

$$\text{Cost} = 100 \text{ bases} \times \$1.95/\text{base} = \$195$$

# Technical considerations

- Synthesis costs

## Custom RNA Synthesis and Purification

IDT has the expertise to deliver custom-synthesized RNA with the yield and purity that today's researcher demands. RNA is shipped deprotected and desalted in 2-3 business days or deprotected and purified in 4-6 business days. Please inquire for turnaround on 5 µmole and 10 µmole RNA synthesis.

Custom RNA Synthesis Pricing:					
	100 nmole	250 nmole	1 µmole	5 µmole	10 µmole
RNA bases	\$6.50 USD	\$8.50 USD	\$18.00 USD	\$60.00 USD	\$115.00 USD

- RNA oligo 100 bases long
- 1 µmol scale

**Cost = 100 bases x \$18/base = \$ 1800**

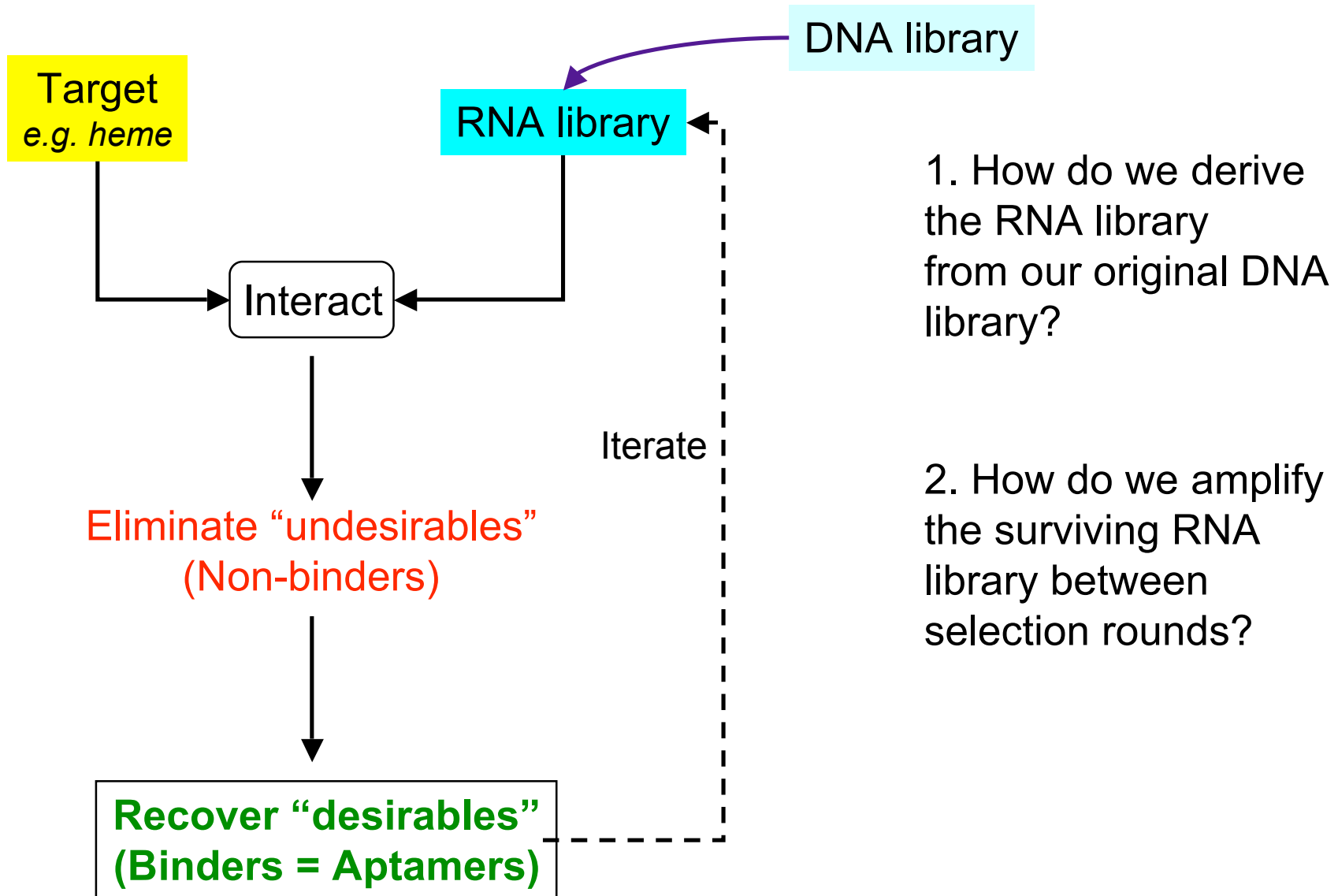


[www.idtdna.com](http://www.idtdna.com)

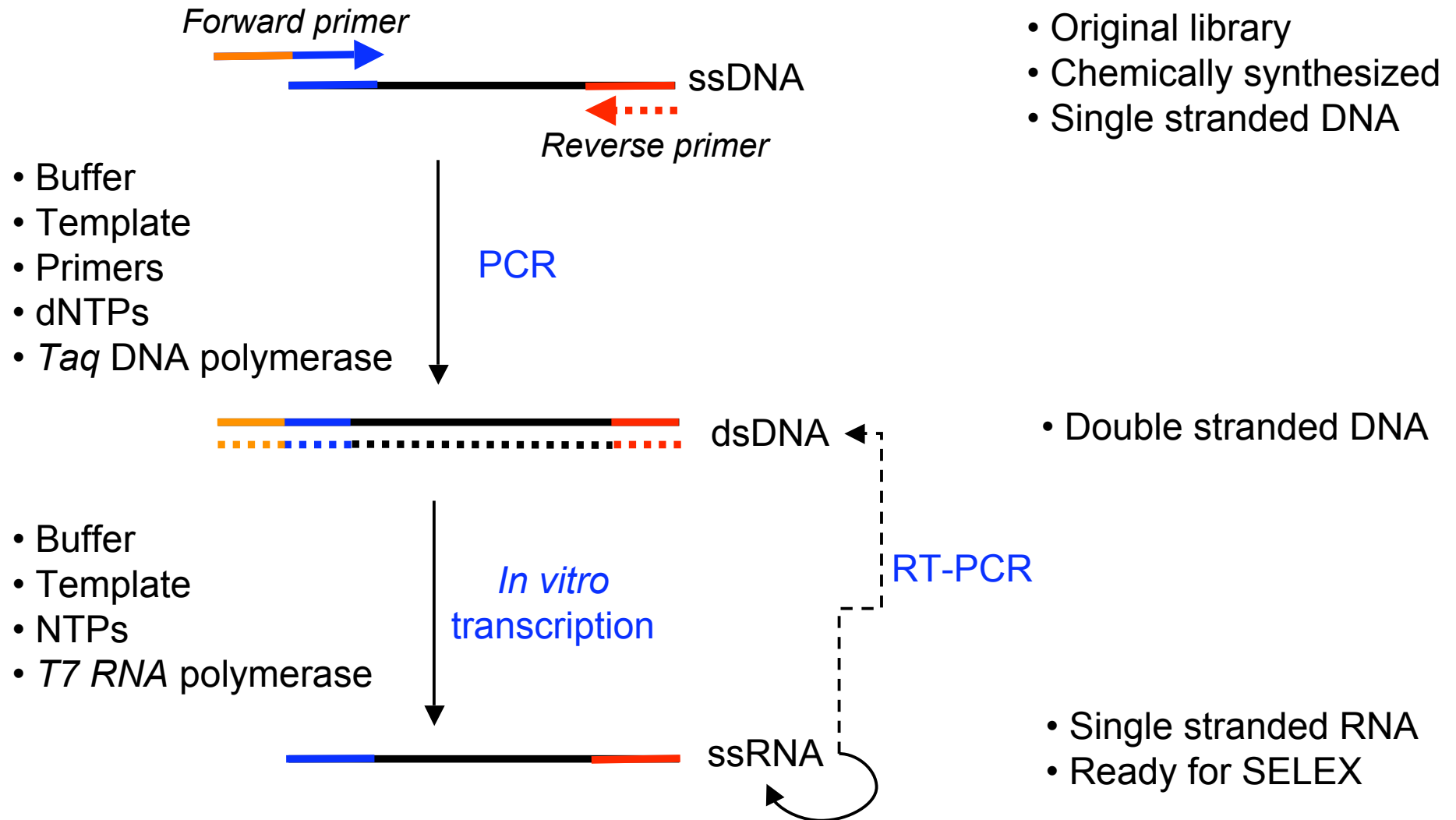
# Technical considerations

- **Stability during storage**
  - *DNA is an excellent long-term form for stably storing library*
- **Cost of synthesis**
  - *DNA is more cost-effective and technically simpler to synthesize than RNA*
- **Two very compelling technical reasons for choosing DNA as the storage medium for your library!**

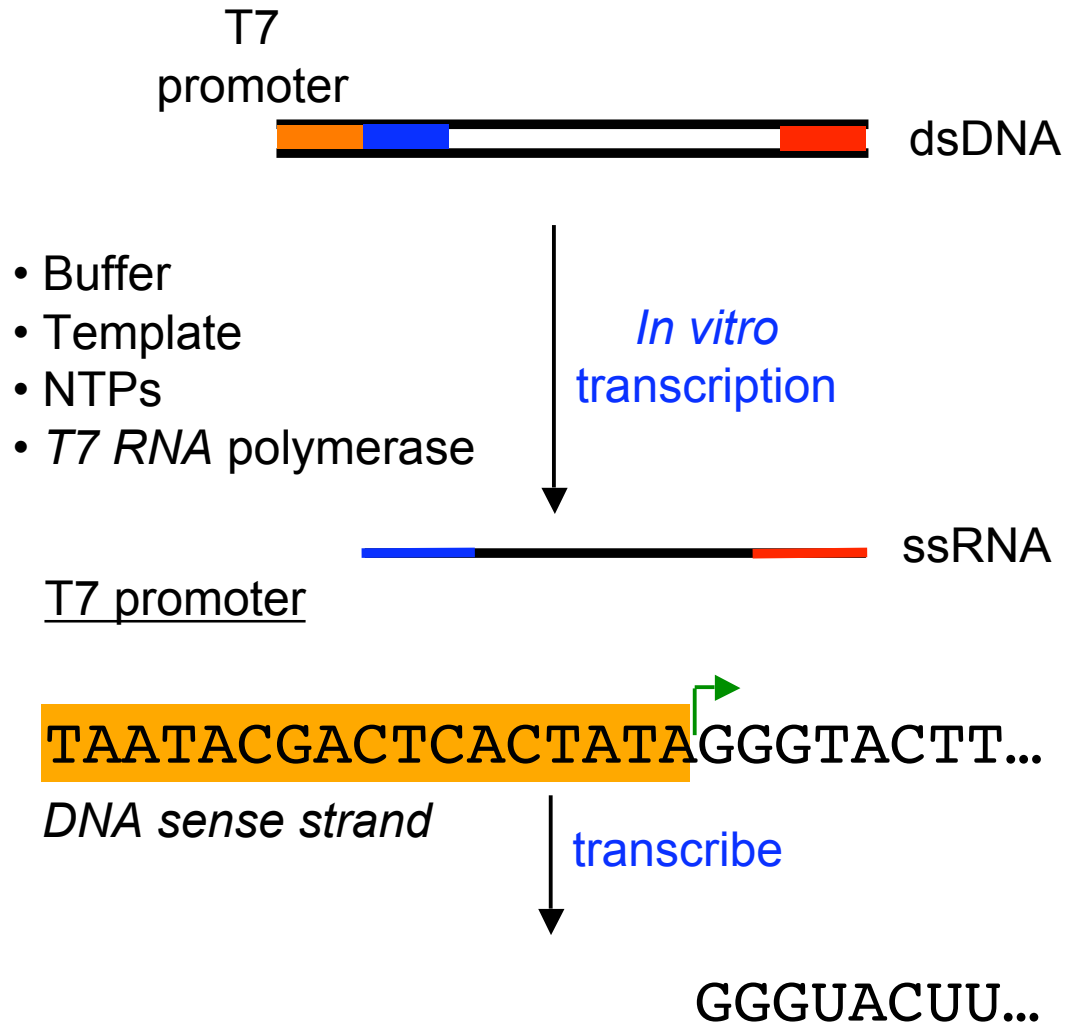
# SELEX: The process (simply)



# SELEX: DNA Library --> RNA Library & Back

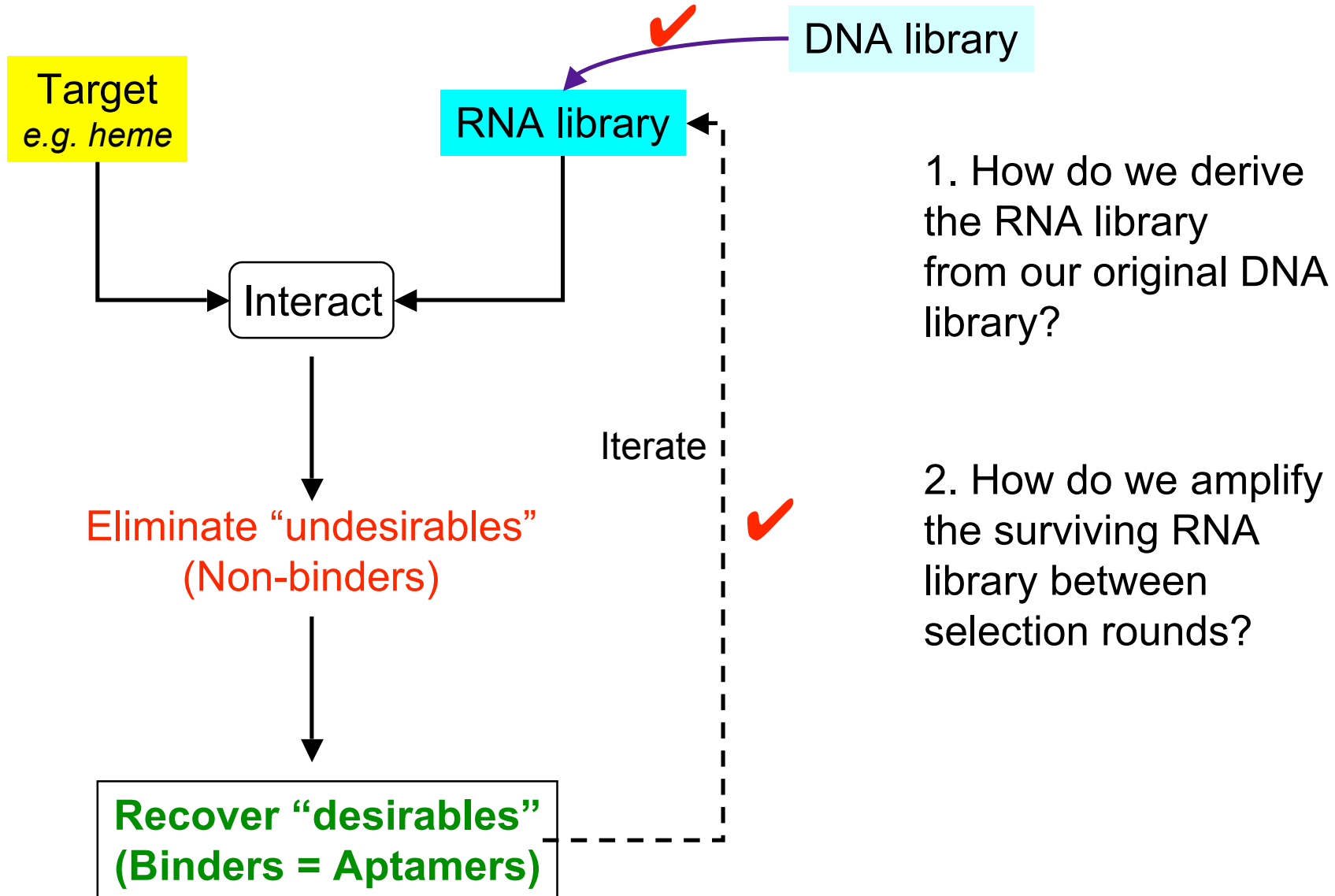


# *In vitro* transcription



T7 RNAP in complex with its promoter  
PDB ([www.rcsb.org](http://www.rcsb.org))

# SELEX: The process (simply)





# Overall architecture of ds DNA library



T7 promoter

**Variable Region**  
(at population level)

– Sequence distinguishes  
one library member from the other!

- Technical constraints dictate this architecture

**How do we achieve variability between individual library members?**



- Each library member has a unique, defined sequence

Member N

- Members differ from each other in the variable region

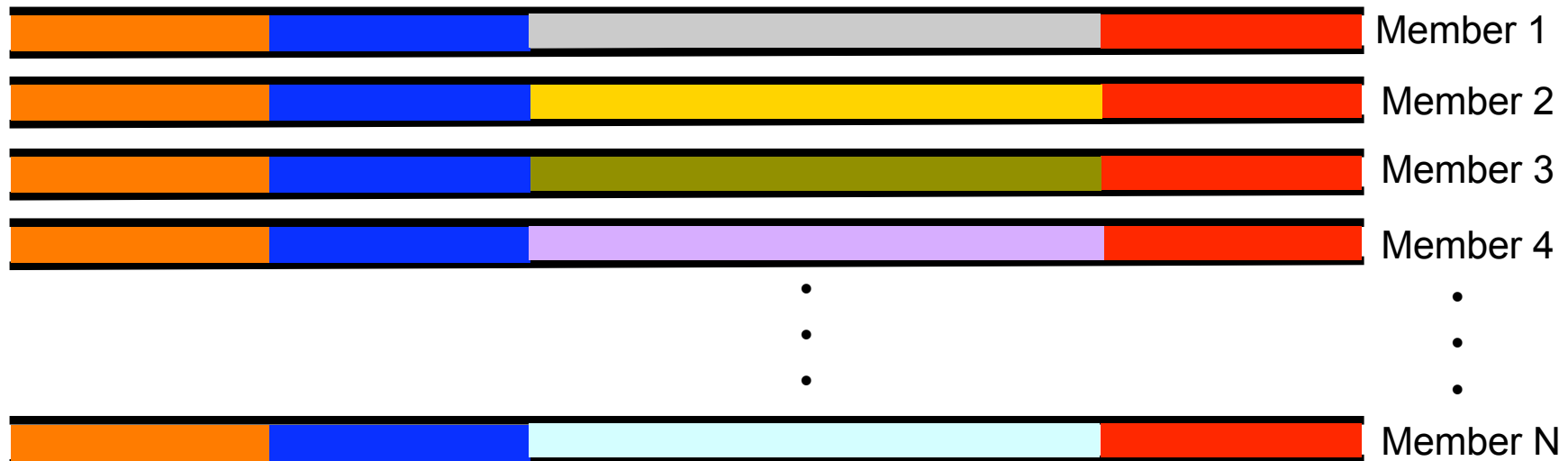
# How do you synthesize such a library?

- DNA synthesis is automated



- Program machine to add a specified base at a specified position

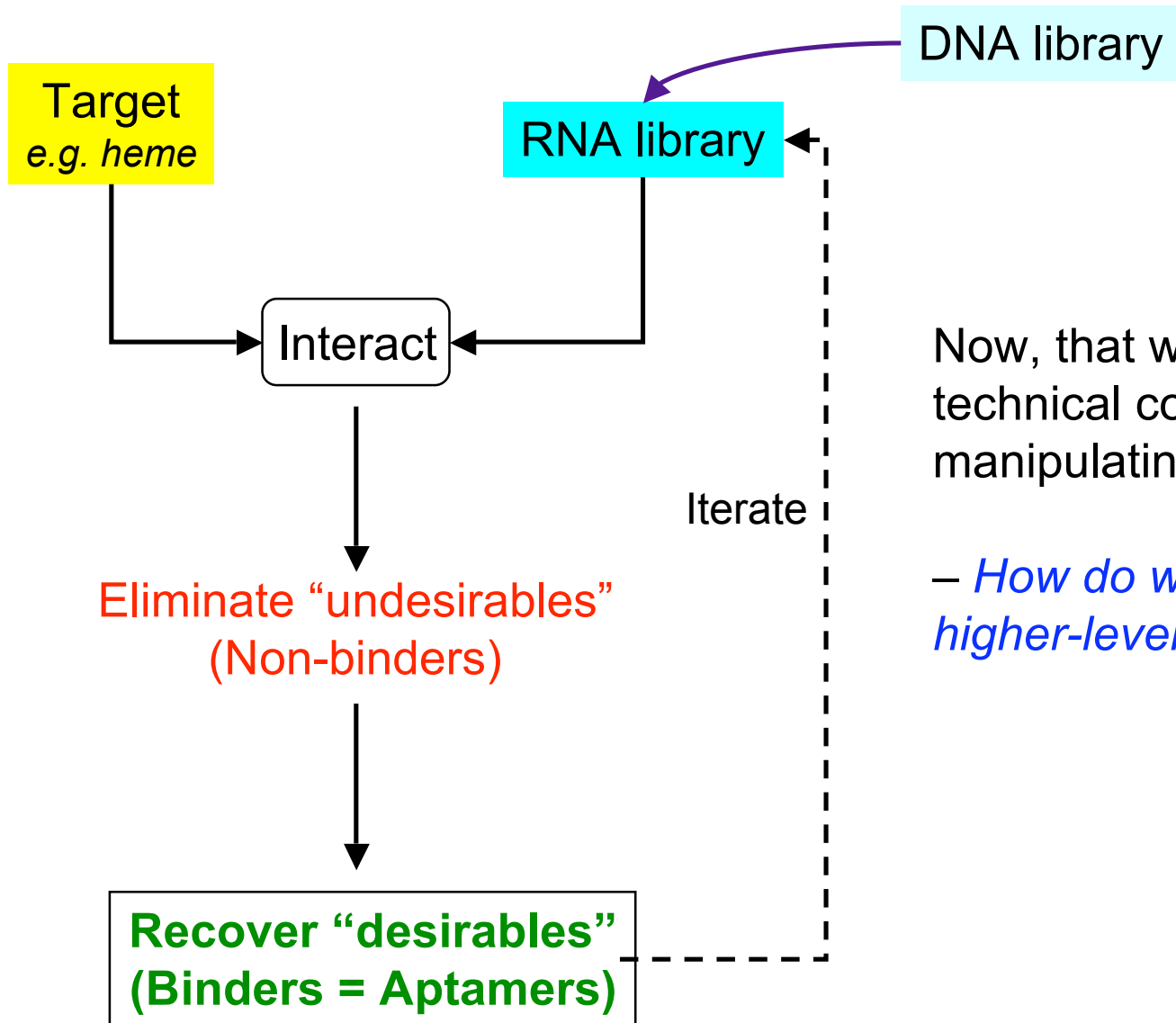
- *How do you build your target library?*



# Exactly as you thought!

- **For fixed regions:**
  - Specify a single nucleotide to be added at that position
- **In the variable region:**
  - Mix the four nucleotides in equal “reactivity” proportions
  - Equal chance of either A, G, T or C being added at that position
  - Many distinct DNA oligonucleotides are being simultaneously synthesized

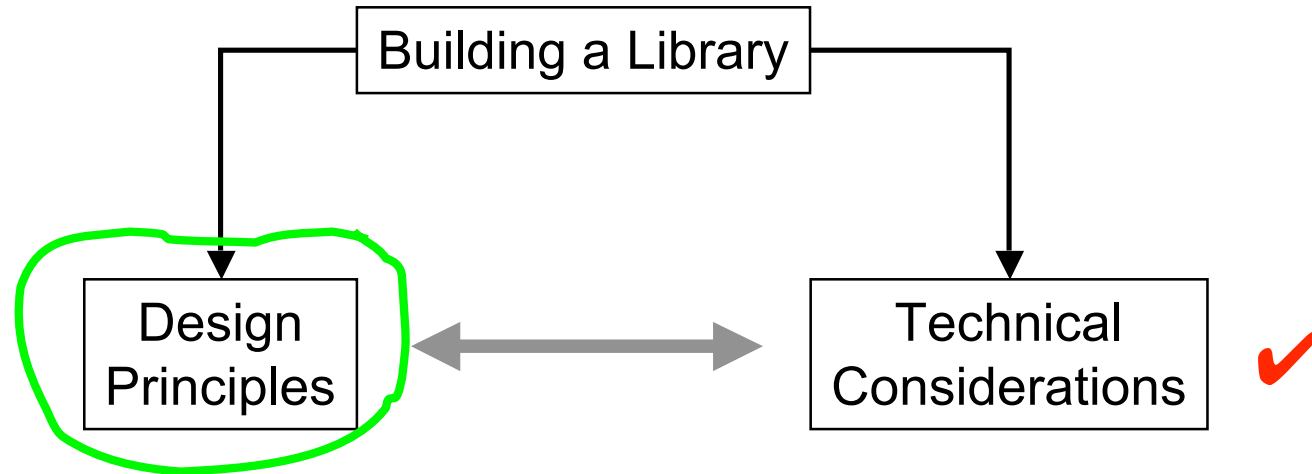
# SELEX: The process (simply)



Now, that we understand the technical constraints for manipulating our library:

– *How do we achieve the higher-level desired properties?*

# The RNA Library (abstracted)



- One library per target **or** *one library for all targets*
- Balance between “useful” and “useless” library members
- Maximizing “useful” collection within space constraints
- *Now, let’s think about what we want in our library!*
- Stability during storage
- Synthesizing library at reasonable costs
- Availability of efficient methods for manipulating library